

Review Article

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Oxidative Stress, Nutrition and Cancer: Friends or Foes?

Salvatore Chirumbolo 

Department of Neurosciences, Biomedicine and Movement Sciences, University of Verona, Verona, Italy

The relationship between cancer and nutrition, as well as nutrition and oxidative stress, shares puzzling aspects that current research is investigating as the possible components of an intriguing regulating mechanism involving the complex interplay between adipose tissue and other compartments. Along the very recent biological evolution, humans underwent a rapid change in their lifestyles and henceforth the role of the adipocytes earned a much more complex task in the fine tuning of the tissue microenvironment. A lipidic signaling language probably evolved in association with the signaling role of reactive oxygen species, which gained a fundamental part in the regulation of cell stem and plasticity. The possible relationship with cancer onset might have some causative mechanism in the impairment of this complex task, usually deregulated by drastic changes in one's own lifestyle and dietary habit. This review tries to address this issue.

Keywords: Adipose tissue; Cancer; Lifestyle; Oxidative stress; Redox

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INTRODUCTION

The great concern of cancer worldwide is expanding its shadows to scientific fields previously far from a forthright relation with oncology. Besides to the awareness that malnutrition and some incorrect dietary habits and lifestyles are possible triggers for cancer onset, current literature is moving its spotlight on the fundamental role exerted by the different adipose tissues in this regards [1-3]. The adipocyte is a highly complex cell, despite its simplistic image of a fat-collecting tank. Current evidence is emerging about the fundamental role of the adipocyte as the master tuner of tissue homeostasis, cross talking with resident immunity and stem cells, where even long non coding RNAs might have a major regulatory commitment [4-7]. In this con-

text, oxidative stress has been long time considered a leading factor in carcinogenesis [8]. Furthermore, the relationship between nutrition intake, dietary habits and different lifestyles with oxidative burden and stress response has been widely and thoroughly addressed so far [9]. The relationship between obesity and cancer might be an interesting vanishing point to focus onto the real function of the different adipose tissues in humans.

A first question may regard which relationship does exist between adipose tissue, metabolism and cancer. This issue is of the utmost importance to shed light on the actual role exerted by different dietary habits on cancer onset and development. The relationship between diabetes and cancer is particularly striking [10,11]. Actually, obese subjects are particularly at risk for

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Correspondence to: Salvatore Chirumbolo  <https://orcid.org/0000-0003-1789-8307>

Department of Neurosciences, Biomedicine and Movement Sciences, University of Verona, Strada Le Grazie 9, 37134 Verona, Italy.

Tel: +39-0458027645, **Fax:** +39-0458027276, **E-mail:** salvatore.chirumbolo@univr.it

cancer etiopathogenesis, though controversial opinions were also forwarded [12-16]. This evidence would suggest that the role of adipocytes in this context is particularly striking. Interestingly, due to their lipid storage, adipocytes might be even considered as stray mines for cancer pathogenesis. Adipocyte-derived lipids may change cancer drug resistance [17] and noteworthy free fatty acids from the adipose tissue are important biomarkers for cancer, as they elicit or even modulate an immuno-inflammatory response [18-20]. Yet, the role of fats in our complex biology and physiology is not so trivial as suspected only few decades ago and therefore even human daily diet to prevent cancer, when addressing fats, should not follow any prejudicial and widespread fashion about fats in the diet, as fats are powerful regulators of the complex cross talk adipose tissue-immunity-tissue turn over. The role of fat molecules, particularly short chain fatty acids, polyunsaturated fatty acids (PUFAs) and more, are currently emerging as fundamental molecules in men's health, as cancer preventive substances [20-24]. The complete, though complex, picture that is emerging from current literature is that the adipocyte is at the cross road of a regulatory network, linking immunity, stress response and cell cycle (with the consequent associated stem mechanisms), where lipids and reactive oxygen species (ROS) act as major signaling molecules. Briefly speaking, the participation of lipids and ROS in creating a master tuner of tissue homeostasis, the leading mechanism that, once disturbed or impaired may cause cancer, is no less fundamental than cytokines and growth factors [25-29]. This overview strongly suggests that lipids intake in the daily diet must be only highly monitored and regulated, not quite completely removed, as the commonest opinion about diet habits would encourage. Particularly for men's health, an unbalanced diet provided with an excess of protein intake, exceeding the Recommended Dietary Assumptions, alongside with poor fats introduction, can be a potential risk for carcinogenesis, although this evidence yet needs to be further assessed [30-33]. For example, a low protein intake, both from animals and plants, in an isocaloric diet, without necessarily following a low-carbohydrate nutritional panel, decreases cancer risk due to a depletion in the CD8+ T cell and antigen presenting cells, mounting an anti-immune response and lowering the inflammatory reaction. This occurrence is related to the triggering of an unfolded protein response in can-

cer cells, a mechanism ignited *via* the activation of the inositol requiring enzyme 1-alpha and retinoic acid inducible gene 1 signaling pathway, resulting in an anti-cancer activity [33].

The role of diet in tuning the adipose tissue function is of the utmost importance. This role is closely associated with further two major components in men's health, namely the gut microbiome brain axis and the musculoskeletal system. The relationship between the skeletal muscle fiber and the adipocyte is emerging in the very recent years, through the role of myokines [34]. The muscle secretome has an important role in modulating the aforementioned role of the adipose tissue in tuning the mechanism of tissue homeostasis, where molecules, such as brain-derived neurotrophic factor and interleukin (IL)-6 directly participate in the AMP-activated protein kinase-mediated lipid oxidation [34]. In this context, even the turn over of ROS is addressed to regulate muscular optimal activity and cell survival, contrarily to the widespread opinion that ROS are deleterious despite their signaling function [35]. Therefore, the synoptic picture we might draw from this perspective is that oxidative stress and nutrients such as fats are collectively bridled to be used as signaling elements for the complex cross talk with tissues that the adipose tissue exerts in its main function.

In this review I am going to deepen and discuss this issue.

A NOVEL HYPOTHESIS ABOUT THE INTRIGUING ROLE OF THE ADIPOSE TISSUE (THE FUNCTIONAL METASTABILITY OF THE ADIPOCYTE)

So far, the role of the adipose tissue, and consequently of the adipocyte as its key cell model, has been ultimately confined to the trivial image of a fat reservoir, with the only fundamental purpose to modelling anatomic regions as a sort of "plastic wax". However, in recent years this commonly widespread view on the adipose tissue has been questioned somehow, by attributing to the adipocyte a major immuno-modulatory role in the organism [36]. Immune function has been long time considered a quite distinct issue respect to bio-energetics, taking it aside from the metabolic and endocrine commitment with which the adipose tissue should be really engaged. Yet, a huge deal of reports

has recently moved the spotlight on the adipocyte as a leading actor in the complex immune-tissues cross talk [37-40]. Despite this, current research did not yet distinguished any purported existence of an exclusive adipocyte-immune cell from the common adipocyte collecting fats in the white adipose tissue (WAT) [36]. On the contrary, the scientific community attributed to the mature adipocyte in WAT the ability to modulate the immune function alongside its main task of lipid storage [41-43]. This allows the adipocyte to be described as having a sort of “secret” double life. Therefore, besides its endocrine role in fat storage, the adipocyte would be an immune-like cell *tout court*, inasmuch the adipose tissue was recently considered a tertiary lymphoid organ [44]. Obviously, this raises many interesting questions about the actual role of the adipose tissue in cancer pathogenesis and development [1,45].

So, is the adipocyte a multi-task cell?

Actually, WAT was reported to home both regulatory T cells (Tregs) and group 2 innate lymphoid cells (ILC2s), keeping the immune function in the tissues under check, a role tuned by IL-33 from the stromal vascular fraction (SVF) [46]. A very recent paper actually showed that lineage negative (lin⁻) SVF cells, which express the platelet-derived growth factor receptor alpha (PDGFR- α) and Sca-1, whereas they lack of the endothelial, hematopoietic and erythroid cell markers CD31, CD45, and Ter111, are active producers of IL-33 [46]. These cells showed typical genotypic features associated with adipose stem and progenitor cells, as expressing *CD34*, *Itgb1* (*CD29*), *Ly6a* (*Sca-1*), *Pdgd*, and *Pdgfra* genes [46]. Although IL-33 has been so far considered a master tuner of innate immunity, its very recent role is to maintain WAT homeostasis [47]. This cytokine is particularly intriguing, as it has been recently related with nutrition, due to the observation that IL-33 regulates the inflammatory process during obesity [48]. Therefore, we are discovering that the adipocyte is not only a fundamental player for the immune homeostasis in WAT but even that immune mediators serve as tuners of the adipose tissue turnover. Conversely, typical adipocyte products, such as lipid droplets, act as immune modulators [49].

A complex immune-adipocyte-tissue plasticity network has inflated the current debate on fat biology also in further men' health concern, such as aesthetic surgery. This still raises these questions: which role the adipocyte is fundamentally committed to, an immune

or a plastic one? Or both? Or have we to turn our attention to a “third” option? Probably, the adipocyte might be considered a sort of sentinel of the “bodily self”, by checking and tuning the correct balance of different tissue types and their dynamics alongside the immune system [50-52].

So, the linkage with stem functions would suggest that the interplay between the adipose tissue and immunity should have a more demanding role than the simple check of a healthy metabolic homeostasis. An immuno-modulatory role, *via* a paracrine pathway, has been very recently recognized to mesenchymal stem cells (MSCs) [53-55], particularly close to the adipocyte biology, so suggesting that the adipocyte might lay at the crossroad immunity/stem-tissue homeostasis, so even suggesting for immunity a wider role within the classical well known immune self. In this perspective, the adipocyte tells of itself as a highly complex cell, finely tuning fundamental and strategic functions in the maintenance of tissue homeostasis, behaving in a much more intriguing way than expected and standing quite far from the sole acknowledged role of a lipid-tank collector. Actually, the very recent reports regarding the adipocyte, would suggest for a new outstanding activity in the “strange” life of the adipocyte, *i.e.*, the adipocyte as a master tuner of tissue homeostasis, not merely a regulator of a more general metabolic steady state. This feature may have important consequences on the comprehension of cancer development.

An intriguing functional kinship with MSCs of hematopoietic origin, sharing apparently a common role in maintaining the biological self *via* immunity, might shed light on the unusual and attractive “existence” of the adipocyte and on the real role of the adipose tissue, which may be of the utmost importance in tissue plasticity [56-58]. The adipocyte life cycle itself should give us insightful clues about the complex activity exerted by the adipose tissue in this perspective.

The first conundrum we should address is whether the white mature adipocyte is a commonly stable, differentiated cell or exhibiting a certain marked tuning ability in its adipose lifecycle. The most widespread view of the differentiated adipocyte is that it is the resulting outcome of the adipogenetic pathway of mesenchymal precursors, usually collectively known as pre-adipocytes. The term pre-adipocytes may be yet outdated, as panoply of adipocyte precursors, called adipocyte progenitor cells (APCs), participate indeed in

the tissue/stem cell homeostasis, which is ultimately ruled by the adipose tissue, as previously suggested [59-62]. The master tuner of pre-adipocyte maturation into adult adipocytes is peroxisome proliferator-activated receptor gamma (PPAR- γ), but further agonists and modulators have been reported to be involved in this mechanism [63,64]. The APC population is formed by cells of mesenchymal origin (MSCs), fibroblast-like cells and a heterogeneous population of cells in the SVF [60,61] of the adipose tissue. Many APCs can be easily identified and isolated [65]. Most of the current research on the adipocyte lifecycle has been carried out on laboratory animals, such as mice or rats. In mice, WAT precursors were identified as PDGFR- α^+ cells with the phenotype CD24⁺, but also the CD24⁻ stem precursors Lin⁻:CD29⁺:CD34⁺:Sca-1⁻:CD24⁻ were identified in the APC population, so suggesting that CD24⁺ cells would generate CD24⁻ cells *in vivo*. CD24⁻ cells will express late markers of the adipogenetic process, *i.e.*, behaving as pre-adipocytes [62]. The loss of CD24 marker may commit APCs to the further adipocyte development [62].

This perspective may shed light on the modulatory role of the adipocyte in tissue homeostasis, encompassing many further issues of men's health, *i.e.*, aging and cancer pathogenesis.

SVF precursors express CD34 (as stem cells) and CD29, whereas did not express peculiar markers of endothelium or hematopoietic lineage, such as CD31, CD45, and Ter111 [62]. Recent reports showed that in mice at least four subgroups of APCs in SVF can be identified on the basis of the expression of CD29 and CD34 but more than half of this population express the stem cell antigen 1 (Sca-1), being Lin⁻:CD29⁺:CD34⁺:Sca-1⁺, being also CD105⁺ and CD117⁺, whereas only a small proportion expressed CD24 [62]. Interestingly, in transgenic mice (A-Zip) lacking of WAT, a dramatic increase in APCs also in the SVF population occurred, with a huge enhancement in the expression of differentiation genes and early markers of adipogenesis, such as Klf4, C/EPB- δ and Krox20, alongside a marked reduction of PPAR- γ [62]. This evidence suggests that SVF contains many stem cells that are not directly committed to become adipocytes unless the occurrence of the expression of defined markers and that a possible feedback circuitry might exist between stem cell precursors and mature adipocytes [66]. Therefore, this complex balance stem precursors/mature adipocytes, if confirmed, should

encompass the ability of mature cells to revert into potential stem lineages. Fig. 1 summarizes one possible model of the complex life cycle of a WAT adipocyte.

In this regard, a fundamental role should be exerted by lipids released by the adipocyte, as signaling molecules in the complex cross talk with other tissues. Actually, mature adipocytes can undergo delipidation. Delipidated adipocytes might be apparently considered aged adipocytes rapidly losing lipids and entering the "senescent" phase of fibroblast-like post-adipocytes but further suggestions were forwarded [67]. Yet, delipidation of mature fat cells was already described in past reports [68]. Delipidation is also associated with the involutive occurrence of terminally-described adipocytes, which appear particularly during sustained stress: the post-adipocytes.

This mechanism makes the adipocyte as a functionally "metastable" cell, in the sense that its stability is not of morphological type but of dynamical one [69].

Furthermore, post-adipocytes have been described in past reports [70] but recently restyled as a mature adipocyte able to resist the massive damage and stress, causing lipid loss, which usually occurs also during lipoaspiration procedures and fat grafting, and to rebuild functional fat cells [67]. Therefore the post-adipocyte is all but a terminal, out of service adipocyte. There might be, therefore, a close relationship between the mechanism of delipidation and the occurrence of post-adipocytes. Delipidated adipocytes suggested in the past the hypothesis that mature most-mitotic adipocytes may not be terminally differentiated, yet [71]. Delipidated adipocyte can give rise *de novo* to mature, fully functional adipocytes [72]. We still do not know if the shift from a post-adipocyte to a mature adipocyte crosses pathways including pre-adipocytes or adipocyte derived stem cells (ADSCs) as precursors of newly adipogenetic cellular pools. To elucidate if post-adipocytes develop into mature adipocyte *via* an ADSC, this cell should express, for example, some specific phenotypic markers, such as CD90, CD44, CD29, CD105, CD13, CD34, CD73, CD166, CD10, CD49e, and CD59 and possibly be negative for markers, such as CD31, CD45, CD14, CD11b, CD19, CD56, and CD146 [73]. Further insights are needed to elucidate this issue.

Furthermore, delipidation may be a highly controlled physiological process, which can occur in ADSCs and leave adipocytes alive, neither undergoing apoptosis or autophagic mechanisms, nor dedifferentiation [74]. For

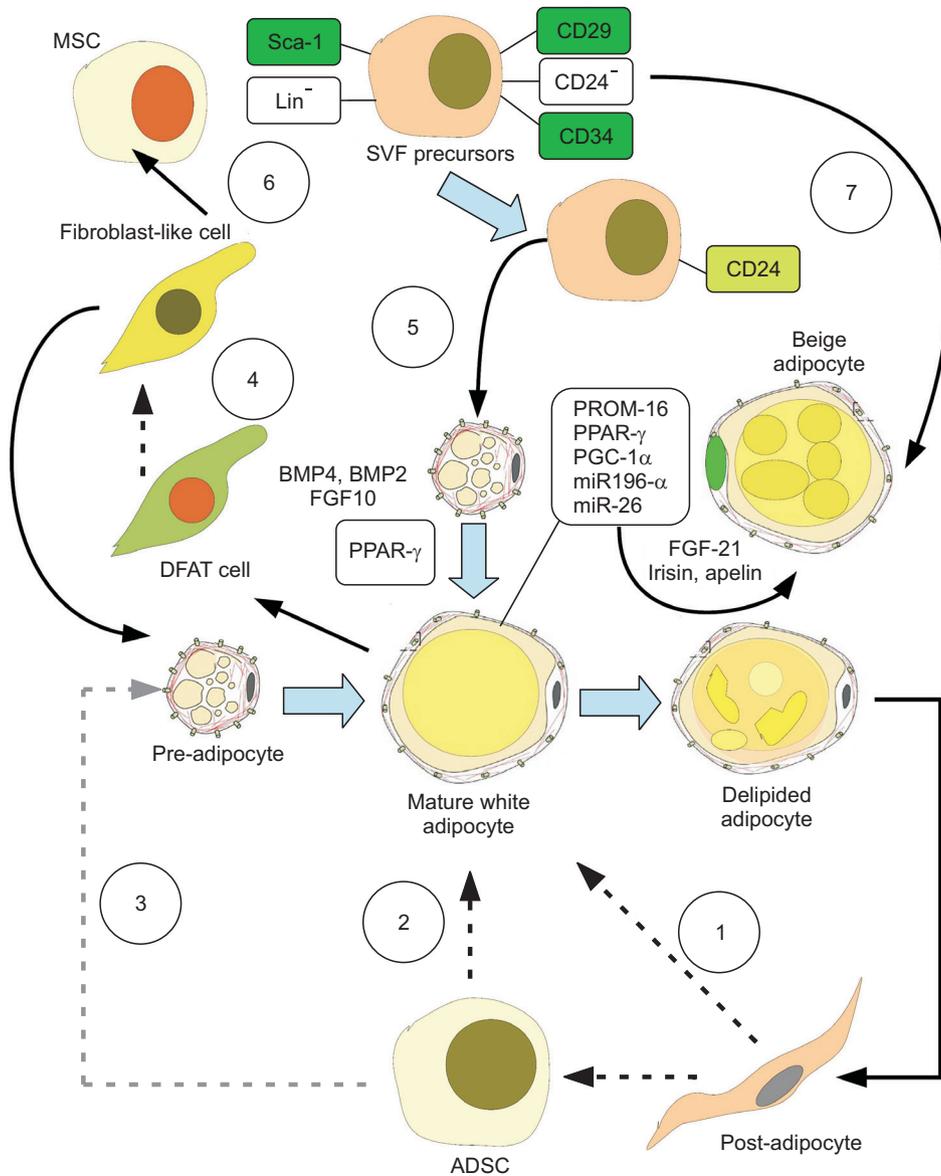


Fig. 1. Hypothesis of the adipocyte life cycle in white adipose tissue. The common routinely pathway is indicated in the centre of the figure (large thick arrows), suggesting the pathway pre-adipocyte-mature adipocyte-delipided adipocyte, which then evolves to a post-adipocyte. This can give rise to adipocytes (1), adipocyte derived stem cells (ADSCs), which then will evolve to adipocytes directly (2) or via a pre-adipocytic pathway (3). Moreover, mature adipocytes can evolve to dedifferentiated adipose tissue (DFAT) cells (4), which might be confused with the stem pool of fibroblast-like cells. In this pool, stromal vascular fraction (SVF) cells are precursors of CD24 non expressing stem cells, which will give rise to white adipocytes (5), while the CD24⁺ SVF pool, which represent the stem cell population together with mesenchymal stem cells (MSCs) (6) will originate beige adipocytes (7). Factors produced by cells (inside the panel) and soluble mediators (out of the panel), which act in this process, are indicated. PPAR: peroxisome proliferator-activated receptor, BMP: bone morphogenetic protein.

example, a controlled nano-induced *in situ* hyperthermia, causes adipocyte delipidation [74]. Interestingly, hypothermia inhibits bone-marrow derived MSCs and increases resistance to hypoxic stress by the SUMOylation of fundamental proteins, such as PCNA, oct-4, p53, and HIF-1α [75]. This evidence suggests that delipidation may be a physiological mechanism of stress response. It is possible to speculate that delipidation

caused by stress, may induce a population of ADSCs which over-expressing heat shock protein-70, enhancing the success and the efficacy of the autologous fat grafting in men's aesthetic health [76]. Further research should address if this population of ADSCs may derive from the delipidation/post-adipocyte route (Fig. 1).

Finally, a dedifferentiation process of the mature adipocyte in WAT, can lead to adipose-derived stem cells

(dedifferentiated adipose tissue, DFAT cells) with fibroblast-like phenotype [77]. There are controversial opinions about DFAT cells, *i.e.*, if these fibroblast-like cells evolved from adipocytes and can reconstitute a new functional adipose tissue (practically post-adipocytes) or if they are true dedifferentiated adipocytes with stem potential [78]. The ambiguity is that fibroblast-like cells inducing new connective cell phenotype may be typical MSCs from the stem pool and not merely DFAT cells [78]. Anyway, this type of cells seem to emerge during a massive loosening of adipose tissue, when stress conditions should induce a fibrotic response [77,78].

THE ROLE OF NUTRITION-DERIVED LIPIDS AND REACTIVE OXYGEN SPECIES

Following the hypothesis suggested in Fig. 1, mature adipocyte in WAT loses lipids in response to stressors, such as ROS and enters a proactive rejuvenation, even passing *via* a stem phenotype. If confirmed, lipids should exert a fundamental signaling role in the mechanism described above about the adipocyte life cycle. Our idea is that adipocytes may “go back” to a relatively younger stage (delipided) and then, depending on the microenvironment conditions, turn again to a mature phenotype or even reaching a stem phenotype, which usually has an adipogenetic commitment. This latest pathway should assure the adipose tissue to enhance its expansion potential. The balance adipocyte precursors/mature adipocytes is fundamental to warrant for a correct anatomical plasticity and tissue homeostasis, a condition that allows to consider the adipocyte a “quasi-stable” or metastable cell, though indicated as a post-mitotic, differentiated cell.

Nutrition has a leading role in this sense.

Recent evidence reported, for example, that dietary conjugated linoleic acid (CLA) reduces the fat burden in adipocytes, *i.e.*, promoted the formation of delipided adipocytes. For example, *trans*-10 and *cis*-12 CLA but not *cis*-9 and *trans*-11 CLA, when added to stromal vascular cells in cell cultures containing newly differentiated human adipocytes, caused time-dependent dampening of the triglyceride content, insulin-stimulated glucose and fatty acid uptake. These same CLAs diminished also PPAR- γ and many PPAR- γ induced genes and increased leptin gene expression. The robust activation of the MEK/ERK signaling preceded

changes in gene expression and caused the release of IL-6 and IL-8. Interestingly, the MEK/ERK activation by *trans*-10 and *cis*-12 CLA was inhibited by UO126 and pertussis toxin [79]. According to these authors, *trans*-10 and *cis*-12 CLA, delipided mature adipocytes by inducing the MEK/ERK signaling *via* an autocrine/paracrine action of adipocytokines, IL-6 and L-8 [79].

It would be particularly interesting to investigate if these lipids are spontaneously formed *in vivo* and, if yes, in which biochemical, physiological (or pathological) stage.

The role of the *trans*-10, *cis*-12 as antagonists of lipidic storage has been widely demonstrated in recent reports [80-83]. This CLA is therefore associated with caloric restriction, anti-obesity molecule, anti-lipid storage factor. Interestingly, this PUFA induces mature adipocyte delipidation [79]. The question is if these lipids are typical signaling mediators of the process in the adipocyte life cycle described before. While the isomer *cis*-9, *trans*-11 induces adipogenesis [84], it is possible to speculate on the existence of cell of “on/off switchers” tuning the ratio t10,c12-CLA/c9,t11 CLA, as they play also a fundamental role in immunity [85]. We yet do not know if adipocytes may have enzymatic switching regulators (as a switch of two double C-C bonds, *cis*-*trans*, shifted of one position towards the ω 6 end with reciprocal isomerization) At least 24 isomers of the 18:2, ω -6 linoleic acid were described [86]. Some studies have investigated the role of CLAs in adipocyte differentiation. The geometric isomer *trans*-10, *cis*-12 CLA reduces the number of pre-adipocytes [87] and inhibits glycerol-3-phosphate dehydrogenase, while *cis*-9 and *trans*-11 antagonized the other [87]. Furthermore, t10,c12 CLA inhibited the expression of PPAR- γ and of sterol regulatory element-binding protein-1c (SREBP-1c) [87] and did not affect the expression of CCAAT/enhancer binding protein alpha (C/EBP- α), therefore t10,c12 CLA cannot be considered a regulator of adipogenesis but of lipid storage and of the balance pre-adipocytes/mature adipocytes, *i.e.*, of delipidation [87,88]. The isomer t10,c12 CLA may induce also inflammation [89]. Moreover, at least in porcine models, CLAs regulates adipogenesis in vascular stromal cells [90]. In these models, t10,c12-CLA inhibits adipogenesis in subcutaneous adipose tissue and promotes adipogenesis in intramuscular adipose tissue [91].

Usually, CLAs are obtained by an α 9 desaturase. In adipocytes PUFAs are regulated by the fatty acid

desaturase FADS1 and FADS2 [91]. In adipocytes linoleic acid is the main tuner of FADS gene expression [91]. Therefore, it is tempting to speculate that CLAs are intrinsic regulators of the adipocyte “biomass” in sub-epidermal layers, where, in this perspective, it is conceivable that fibroblasts and keratinocytes may be fundamental sources of linoleic acid. Linoleic acid, in the presence of β -NADH, elicits a rapid and immediate ROS production in fibroblasts, which most probably act as signaling molecules to activate fibroblast oxidative burst [92].

It is also well known that ROS induce adipocyte differentiation *via* an increased expression of PPAR- γ and activating the CCAAT/enhancer-binding protein β (C/EBP- β) [93]. Anyway, this adipocyte differentiation occurs, at least in culture, if cells reach the confluence and under ROS oxidation. In this conditions, in the constant presence of inducing hormones, adipocytes activate PPAR- γ and differentiate into mature adipocytes and ROS enhanced cell phase S during the mitotic clonal expansion [93]. This is quite probably, the commonest, most orthodox circumstance regarding adiposity modulation in sub-epidermal fat tissues. Detached adipocytes are more likely to loss lipids and to enter a route of dedifferentiation, while adipocytes still present in a whole adipose tissue respond to the aforementioned cycle.

Conversely, ROS might activate also dedifferentiated adipocytes, most probably *via* a Notch/NADPH oxidase 4 interplay, but usually when ROS exceed a certain threshold, probably evaluated by mitochondria oscillation [94-96]. According to this hypothesis, there must exist a lipid signaling as a sort of “warning” signal, able to induce the adipocyte to shifting towards “younger” phenotypes and so enabling the adipose tissue to respond to the stressful stimulus maintaining the tissue homeostasis of the organism. Much ADSCs and dedifferentiated adipocytes might be therefore elicited when a defined “threshold”, probably induced by a certain milieu of soluble factors and cytokines, is overwhelmed, usually quite far from physiological conditions. Further research should yet elucidate this issue.

The role of ROS is therefore fundamental as a signaling pathway to set the adipocyte at the crossroad of the complex interplay energetic metabolism/tissue homeostasis. Therefore, they have a fundamental action also in the evolution of cancer, taking into account

these issues.

The extreme plasticity of the adipocyte is a leading cause of many tumors, due to the occurring impairment in this complex cross talk [97].

In this perspective, the different type of adipose tissues in humans and the different types of fat composition in adipocytes, should exert a fundamental role in tissue homeostasis, so driving the complex signaling machinery preventing cancer in the organism.

Fats are therefore signaling molecules as well as ROS, a perspective that should raise the question if fats and ROS are more friends than foes for men's health. A first interesting evidence is the role of lipids stored in cells as lipid droplets to counteract oxidative stress and move onward the metabolic-energetic machinery of cells [98]. Actually, lipid droplets are able to tune cell oxidative stress [99]. According to some authors, lipid droplets have the main property to buffer and modulate the delayed release of fats, both as signaling molecules and metabolic sources, and the delayed release is fundamental for many purposes, such as energy and redox homeostasis, tuning of autophagy [100] and of the ER stress [99]. In this sense, lipids actively participate as mediators in the cross talk between cells and tissues. Lipids in diet are therefore fundamental as possible elements in the creation of a complex signaling language among cells and within cells [101].

In this landscape, ROS may have also a fundamental task.

PPAR- γ and autophagy are intertwined in controlling both ROS signaling and lipid droplets production and turn over, having therefore a role in some type of cancer, such as colon cancer [102]. Actually, the role of PPAR- γ is more intriguing than expected, as it may tune the so called “two-compartment tumor metabolism” model, where a metabolic coupling has been suggested to occur between catabolic stromal cells and oxidative tumor cells [103]. In this models, cancers, for example breast cancer, is embedded in a complex microenvironment where a cross talk cancer cells/stromal cells occurs, in order to tune cancer survival. This cross talk involves also adipocytes. Cancer cells elicit in stromal cells glycolysis, lipid droplets, ROS formation from mitochondria and autophagy, while stromal cells activate a response with onco-metabolites that include fatty acids and peroxidized lipids [103].

Lipids modified by ROS and activated oxygen, are

powerful signal molecules even in plants [104].

For example, oxysterols are fundamental signaling molecules in cancer [105,106]. These molecules, occurring as cholesterol oxidized metabolites, may be produced by radical (ROS-dependent) mechanisms and make the so called oxysterome. They are fundamental actors in the tissue turnover, including cancer etiology, by interacting with a huge panoply of receptors, such as nuclear receptors (ROR, ER α , LXR) as well as protein G coupled receptors (CXCR2, SMO, EBI2, and so on) and becoming fundamental biomarkers in many chronic pathologies, including cancer, such as 4 β -hydroxy-cholesterol or 7 α -hydroxy-cholestenone [107].

The very recent awareness in science that lipids and their metabolites, together with lipid droplets, participate in the complex signaling language between tissues, so having a fundamental role in cancer, have put adipose tissue in a strategic focus, an intriguing perspective, particularly because adipocytes are able to develop stem features. Therefore, the new engagement in the biomedical research to highlight any aspects improving men's health, is to take into consideration this fundamental issue.

CONCLUSIONS

This review would simply address some fundamental "hot topics" that deserve further attention and interest in science to attain a real improvement in our knowledge on men's health and the onset of cancer. A huge deal of data regarding cancer and nutrition exists elsewhere but this manuscript aimed at stressing the major items able to drive further investigation towards this target, with a simple recommendation. Preventing diets from lipids at all, may not be a wise counselling to prevent chronic pathologies and cancer. Moderation is always advised but lipids are fundamental actors of our own health, as they can act as modulatory signaling molecules. Even ROS, in his perspective, may have a positive role in promoting health.

Therefore further research is needed to elucidate the complex role of ROS and adipose tissue in the different dietary habits engaged by humans.

Conflicts of Interest

The author has nothing to disclose.

Author Contribution

Conceptualization: SC. Data curation: SC. Formal analysis SC. Investigation: SC. Methodology: SC. Project administration: SC. Resources: SC. Software: SC. Supervision: SC. Validation: SC. Visualization: SC. Writing – original draft: SC. Writing – review & editing: SC.

REFERENCES

1. Zimta AA, Tigu AB, Muntean M, Cenariu D, Slaby O, Berindan-Neagoe I. Molecular links between central obesity and breast cancer. *Int J Mol Sci* 2019;20:E5364.
2. Agurs-Collins T, Ross SA, Dunn BK. The many faces of obesity and its influence on breast cancer risk. *Front Oncol* 2019; 9:765.
3. Himbert C, Delphan M, Scherer D, Bowers LW, Hursting S, Ulrich CM. Signals from the adipose microenvironment and the obesity-cancer link: a systematic review. *Cancer Prev Res (Phila)* 2017;10:494-506.
4. Chen K, Xie S, Jin W. Crucial lncRNAs associated with adipocyte differentiation from human adipose-derived stem cells based on co-expression and ceRNA network analyses. *PeerJ* 2019;7:e7544.
5. Kopp A, Buechler C, Neumeier M, Weigert J, Aslanidis C, Schölmerich J, et al. Innate immunity and adipocyte function: ligand-specific activation of multiple Toll-like receptors modulates cytokine, adipokine, and chemokine secretion in adipocytes. *Obesity (Silver Spring)* 2009;17:648-56.
6. Wu Q, Li B, Li Z, Li J, Sun S, Sun S. Cancer-associated adipocytes: key players in breast cancer progression. *J Hematol Oncol* 2019;12:95.
7. Chen SX, Zhang LJ, Gallo RL. Dermal white adipose tissue: a newly recognized layer of skin innate defense. *J Invest Dermatol* 2019;139:1002-9.
8. Klaunig JE, Wang Z. Oxidative stress in carcinogenesis. *Curr Opin Toxicol* 2018;7:116-21.
9. Björklund G, Chirumbolo S. Role of oxidative stress and antioxidants in daily nutrition and human health. *Nutrition* 2017; 33:311-21.
10. Giovannucci E, Harlan DM, Archer MC, Bergenstal RM, Gapstur SM, Habel LA, et al. Diabetes and cancer: a consensus report. *Diabetes Care* 2010;33:1674-85.
11. Collins KK. The diabetes-cancer link. *Diabetes Spectr* 2014; 27:276-80.
12. Salaün H, Thariat J, Vignot M, Merrouche Y, Vignot S. [Obesity and cancer]. *Bull Cancer* 2017;104:30-41. French.
13. Lennon H, Sperrin M, Badrick E, Renehan AG. The obesity

- paradox in cancer: a review. *Curr Oncol Rep* 2016;18:56.
14. Iyengar NM, Gucalp A, Dannenberg AJ, Hudis CA. Obesity and cancer mechanisms: tumor microenvironment and inflammation. *J Clin Oncol* 2016;34:4270-6.
 15. Nindrea RD, Aryandono T, Lazuardi L, Dwiprahasto I. Association of overweight and obesity with breast cancer during premenopausal period in Asia: a meta-analysis. *Int J Prev Med* 2019;10:192.
 16. Aurilio G, Piva F, Santoni M, Cimadamore A, Sorgentoni G, Lopez-Beltran A, et al. The role of obesity in renal cell carcinoma patients: clinical-pathological implications. *Int J Mol Sci* 2019;20:E5683.
 17. Cao Y. Adipocyte and lipid metabolism in cancer drug resistance. *J Clin Invest* 2019;129:3006-17.
 18. Hopkins MM, Meier KE. Free fatty acid receptors and cancer: from nutrition to pharmacology. *Handb Exp Pharmacol* 2017; 236:233-51.
 19. Liu J, Mazzone PJ, Cata JP, Kurz A, Bauer M, Mascha EJ, et al. Serum free fatty acid biomarkers of lung cancer. *Chest* 2014; 146:670-9.
 20. Yeop Han C, Kargi AY, Omer M, Chan CK, Wabitsch M, O'Brien KD, et al. Differential effect of saturated and unsaturated free fatty acids on the generation of monocyte adhesion and chemotactic factors by adipocytes: dissociation of adipocyte hypertrophy from inflammation. *Diabetes* 2010;59:386-96.
 21. Sivaprakasam S, Prasad PD, Singh N. Benefits of short-chain fatty acids and their receptors in inflammation and carcinogenesis. *Pharmacol Ther* 2016;164:144-51.
 22. Thirunavukkarasan M, Wang C, Rao A, Hind T, Teo YR, Siddiquee AA, et al. Short-chain fatty acid receptors inhibit invasive phenotypes in breast cancer cells. *PLoS One* 2017;12: e0186334.
 23. Abel S, Riedel S, Gelderblom WC. Dietary PUFA and cancer. *Proc Nutr Soc* 2014;73:361-7.
 24. Serini S, Ottens Vasconcelos R, Fasano E, Calviello G. How plausible is the use of dietary n-3 PUFA in the adjuvant therapy of cancer? *Nutr Res Rev* 2016;29:102-25.
 25. Ray U, Roy SS. Aberrant lipid metabolism in cancer cells - the role of oncolipid-activated signaling. *FEBS J* 2018;285:432-43.
 26. Peck B, Schulze A. Lipid desaturation - the next step in targeting lipogenesis in cancer? *FEBS J* 2016;283:2767-78.
 27. Wang P, Yuan Y, Lin W, Zhong H, Xu K, Qi X. Roles of sphingosine-1-phosphate signaling in cancer. *Cancer Cell Int* 2019; 19:295.
 28. Kallunki T, Olsen OD, Jäättelä M. Cancer-associated lysosomal changes: friends or foes? *Oncogene* 2013;32:1995-2004.
 29. Wu L, Tang Q, Yin X, Yan D, Tang M, Xin J, et al. The therapeutic potential of adipose tissue-derived mesenchymal stem cells to enhance radiotherapy effects on hepatocellular carcinoma. *Front Cell Dev Biol* 2019;7:267.
 30. Delimaris I. Adverse effects associated with protein intake above the recommended dietary allowance for adults. *ISRN Nutr* 2013;2013:126929.
 31. Lai R, Bian Z, Lin H, Ren J, Zhou H, Guo H. The association between dietary protein intake and colorectal cancer risk: a meta-analysis. *World J Surg Oncol* 2017;15:169.
 32. Pili R, Fontana L. Low-protein diet in cancer: ready for prime time? *Nat Rev Endocrinol* 2018;14:384-6.
 33. Rubio-Patiño C, Bossowski JP, De Donatis GM, Mondragón L, Villa E, Aira LE, et al. Low-protein diet induces IRE1 α -dependent anticancer immunosurveillance. *Cell Metab* 2018; 27:828-42.e7.
 34. Pedersen BK, Febbraio MA. Muscles, exercise and obesity: skeletal muscle as a secretory organ. *Nat Rev Endocrinol* 2012;8:457-65.
 35. Steinbacher P, Eckl P. Impact of oxidative stress on exercising skeletal muscle. *Biomolecules* 2015;5:356-77.
 36. Corrêa LH, Heyn GS, Magalhaes KG. The impact of the adipose organ plasticity on inflammation and cancer progression. *Cells* 2019;8:E662.
 37. Engin AB. Adipocyte-macrophage cross-talk in obesity. *Adv Exp Med Biol* 2017;960:327-43.
 38. Wong Y, Nakamizo S, Tan KJ, Kabashima K. An update on the role of adipose tissues in psoriasis. *Front Immunol* 2019;10: 1507.
 39. Zhou H, Liu F. Regulation, communication, and functional roles of adipose tissue-resident CD4⁺ T cells in the control of metabolic homeostasis. *Front Immunol* 2018;9:1961.
 40. Kumari M, Heeren J, Scheja L. Regulation of immunometabolism in adipose tissue. *Semin Immunopathol* 2018;40:189-202.
 41. Wensveen FM, Valentić S, Šestan M, Wensveen TT, Polić B. Interactions between adipose tissue and the immune system in health and malnutrition. *Semin Immunol* 2015;27:322-33.
 42. Apostolopoulos V, de Courten MP, Stojanovska L, Blatch GL, Tangalakis K, de Courten B. The complex immunological and inflammatory network of adipose tissue in obesity. *Mol Nutr Food Res* 2016;60:43-57.
 43. Meijer K, de Vries M, Al-Lahham S, Bruinenberg M, Weening D, Dijkstra M, et al. Human primary adipocytes exhibit immune cell function: adipocytes prime inflammation independent of macrophages. *PLoS One* 2011;6:e17154.
 44. Frasca D, Blomberg BB. Adipose tissue: a tertiary lymphoid organ: does it change with age? *Gerontology* 2019. doi: 10.1159/000502036 [Epub].

45. Bielczyk-Maczynska E. White adipocyte plasticity in physiology and disease. *Cells* 2019;8:E1507.
46. Mahlaköiv T, Flamar AL, Johnston LK, Moriyama S, Putzel GG, Bryce PJ, et al. Stromal cells maintain immune cell homeostasis in adipose tissue via production of interleukin-33. *Sci Immunol* 2019;4:eaax0416.
47. Dempsey LA. Fat IL-33 sources. *Nat Immunol* 2019;20:776.
48. de Oliveira MFA, Talvani A, Rocha-Vieira E. IL-33 in obesity: where do we go from here? *Inflamm Res* 2019;68:185-94.
49. den Brok MH, Raaijmakers TK, Collado-Camps E, Adema GJ. Lipid droplets as immune modulators in myeloid cells. *Trends Immunol* 2018;39:380-92.
50. Dierickx P, Van Laake LW, Geijsen N. Circadian clocks: from stem cells to tissue homeostasis and regeneration. *EMBO Rep* 2018;19:18-28.
51. Janich P, Meng QJ, Benitah SA. Circadian control of tissue homeostasis and adult stem cells. *Curr Opin Cell Biol* 2014;31:8-15.
52. Ninel Hansen S, Peics J, Gerhart-Hines Z. Keeping fat on time: circadian control of adipose tissue. *Exp Cell Res* 2017;360:31-4.
53. Zhou Y, Yamamoto Y, Xiao Z, Ochiya T. The immunomodulatory functions of mesenchymal stromal/stem cells mediated via paracrine activity. *J Clin Med* 2019;8:E1025.
54. Gruber HE, Deepe R, Hoelscher GL, Ingram JA, Norton HJ, Scannell B, et al. Human adipose-derived mesenchymal stem cells: direction to a phenotype sharing similarities with the disc, gene expression profiling, and coculture with human annulus cells. *Tissue Eng Part A* 2010;16:2843-60.
55. Salami F, Tavassoli A, Mehrzad J, Parham A. Immunomodulatory effects of mesenchymal stem cells on leukocytes with emphasis on neutrophils. *Immunobiology* 2018;223:786-91.
56. Baer PC. Adipose-derived mesenchymal stromal/stem cells: an update on their phenotype in vivo and in vitro. *World J Stem Cells* 2014;6:256-65.
57. Baer PC, Geiger H. Adipose-derived mesenchymal stromal/stem cells: tissue localization, characterization, and heterogeneity. *Stem Cells Int* 2012;2012:812693.
58. Kolaparthi LK, Sanivarapu S, Moogla S, Kutcham RS. Adipose tissue - adequate, accessible regenerative material. *Int J Stem Cells* 2015;8:121-7.
59. Rosen ED, Spiegelman BM. What we talk about when we talk about fat. *Cell* 2014;156:20-44.
60. Hepler C, Vishvanath L, Gupta RK. Sorting out adipocyte precursors and their role in physiology and disease. *Genes Dev* 2017;31:127-40.
61. Rodeheffer MS, Birsoy K, Friedman JM. Identification of white adipocyte progenitor cells in vivo. *Cell* 2008;135:240-9.
62. Berry R, Rodeheffer MS. Characterization of the adipocyte cellular lineage in vivo. *Nat Cell Biol* 2013;15:302-8.
63. Farmer SR. Transcriptional control of adipocyte formation. *Cell Metab* 2006;4:263-73.
64. Cristancho AG, Lazar MA. Forming functional fat: a growing understanding of adipocyte differentiation. *Nat Rev Mol Cell Biol* 2011;12:722-34.
65. Church CD, Berry R, Rodeheffer MS. Isolation and study of adipocyte precursors. *Methods Enzymol* 2014;537:31-46.
66. Spalding KL, Arner E, Westermark PO, Bernard S, Buchholz BA, Bergmann O, et al. Dynamics of fat cell turnover in humans. *Nature* 2008;453:783-7.
67. Conti G, Benati D, Bernardi P, Jurga M, Rigotti G, Sbarbati A. The post-adipocytic phase of the adipose cell cycle. *Tissue Cell* 2014;46:520-6.
68. Petruschke T, Hauner H. Tumor necrosis factor-alpha prevents the differentiation of human adipocyte precursor cells and causes delipidation of newly developed fat cells. *J Clin Endocrinol Metab* 1993;76:742-7.
69. Rigotti G, Chirumbolo S. Biological morphogenetic surgery: a minimally invasive procedure to address different biological mechanisms. *Aesthet Surg J* 2019;39:745-55.
70. Unger RH, Zhou YT, Orci L. Regulation of fatty acid homeostasis in cells: novel role of leptin. *Proc Natl Acad Sci U S A* 1999;96:2327-32.
71. Cryer A, Van RLR. *New perspectives in adipose tissue: structure, function and development*. London: Butterworths; 1985.
72. Tavassoli M. In vivo development of adipose tissue following implantation of lipid-depleted cultured adipocyte. *Exp Cell Res* 1982;137:55-62.
73. Mildmay-White A, Khan W. Cell surface markers on adipose-derived stem cells: a systematic review. *Curr Stem Cell Res Ther* 2017;12:484-92.
74. Marinozzi MR, Pandolfi L, Malatesta M, Colombo M, Collico V, Lievens PM, et al. Innovative approach to safely induce controlled lipolysis by superparamagnetic iron oxide nanoparticles-mediated hyperthermic treatment. *Int J Biochem Cell Biol* 2017;93:62-73.
75. Liu X, Ren W, Jiang Z, Su Z, Ma X, Li Y, et al. Hypothermia inhibits the proliferation of bone marrow-derived mesenchymal stem cells and increases tolerance to hypoxia by enhancing SUMOylation. *Int J Mol Med* 2017;40:1631-8.
76. Feng H, Qiu L, Zhang T, Yu H, Ma X, Su Y, et al. Heat-shock protein 70 overexpression in adipose-derived stem cells enhances fat graft survival. *Ann Plast Surg* 2017;78:460-6.
77. Chirumbolo S, Bjørklund G. Can Wnt5a and Wnt non-canonical pathways really mediate adipocyte de-differentiation in a tumour microenvironment? *Eur J Cancer* 2016;64:96-

- 100.
78. Liao Y, Zeng Z, Lu F, Dong Z, Chang Q, Gao J. In vivo de-differentiation of adult adipose cells. *PLoS One* 2015;10: e0125254.
 79. Brown JM, Boysen MS, Chung S, Fabiyi O, Morrison RF, Mandrup S, et al. Conjugated linoleic acid induces human adipocyte delipidation: autocrine/paracrine regulation of MEK/ERK signaling by adipocytokines. *J Biol Chem* 2004;279: 26735-47.
 80. Obsen T, Faergeman NJ, Chung S, Martinez K, Govern S, Loreau O, et al. Trans-10, cis-12 conjugated linoleic acid decreases de novo lipid synthesis in human adipocytes. *J Nutr Biochem* 2012;23:580-90.
 81. Marques TM, Wall R, O'Sullivan O, Fitzgerald GF, Shanahan F, Quigley EM, et al. Dietary trans-10, cis-12-conjugated linoleic acid alters fatty acid metabolism and microbiota composition in mice. *Br J Nutr* 2015;113:728-38.
 82. Yeganeh A, Zahradka P, Taylor CG. Trans-10,cis-12 conjugated linoleic acid (t10-c12 CLA) treatment and caloric restriction differentially affect adipocyte cell turnover in obese and lean mice. *J Nutr Biochem* 2017;49:123-32.
 83. Kennedy A, Chung S, LaPoint K, Fabiyi O, McIntosh MK. Trans-10, cis-12 conjugated linoleic acid antagonizes ligand-dependent PPAR γ activity in primary cultures of human adipocytes. *J Nutr* 2008;138:455-61.
 84. Segovia SA, Vickers MH, Gray C, Zhang XD, Reynolds CM. Conjugated linoleic acid supplementation improves maternal high fat diet-induced programming of metabolic dysfunction in adult male rat offspring. *Sci Rep* 2017;7:6663.
 85. Albers R, van der Wielen RP, Brink EJ, Hendriks HF, Dorowska-Taran VN, Mohede IC. Effects of cis-9, trans-11 and trans-10, cis-12 conjugated linoleic acid (CLA) isomers on immune function in healthy men. *Eur J Clin Nutr* 2003;57: 595-603.
 86. Banni S. Conjugated linoleic acid metabolism. *Curr Opin Lipidol* 2002;13:261-6.
 87. Brandebourg TD, Hu CY. Isomer-specific regulation of differentiating pig preadipocytes by conjugated linoleic acids. *J Anim Sci* 2005;83:2096-105.
 88. House RL, Cassady JP, Eisen EJ, McIntosh MK, Odle J. Conjugated linoleic acid evokes de-lipidation through the regulation of genes controlling lipid metabolism in adipose and liver tissue. *Obes Rev* 2005;6:247-58.
 89. LaRosa PC, Miner J, Xia Y, Zhou Y, Kachman S, Fromm ME. Trans-10, cis-12 conjugated linoleic acid causes inflammation and delipidation of white adipose tissue in mice: a microarray and histological analysis. *Physiol Genomics* 2006;27:282-94.
 90. Zhou X, Li D, Yin J, Ni J, Dong B, Zhang J, et al. CLA differently regulates adipogenesis in stromal vascular cells from porcine subcutaneous adipose and skeletal muscle. *J Lipid Res* 2007;48:1701-9.
 91. Ralston JC, Matravada S, Gaudio N, Holloway GP, Mutch DM. Polyunsaturated fatty acid regulation of adipocyte FADS1 and FADS2 expression and function. *Obesity (Silver Spring)* 2015;23:725-8.
 92. Hatanaka E, Dermargos A, Hirata AE, Vinolo MA, Carpinelli AR, Newsholme P, et al. Oleic, linoleic and linolenic acids increase ros production by fibroblasts via NADPH oxidase activation. *PLoS One* 2013;8:e58626.
 93. Lee H, Lee YJ, Choi H, Ko EH, Kim JW. Reactive oxygen species facilitate adipocyte differentiation by accelerating mitotic clonal expansion. *J Biol Chem* 2009;284:10601-9.
 94. Jiao W, Ji J, Li F, Guo J, Zheng Y, Li S, et al. Activation of the Notch-Nox4-reactive oxygen species signaling pathway induces cell death in high glucose-treated human retinal endothelial cells. *Mol Med Rep* 2019;19:667-77.
 95. Chirumbolo S, Björklund G. PERM hypothesis: the fundamental machinery able to elucidate the role of xenobiotics and hormesis in cell survival and homeostasis. *Int J Mol Sci* 2017;18:E165.
 96. Castro JP, Grune T, Speckmann B. The two faces of reactive oxygen species (ROS) in adipocyte function and dysfunction. *Biol Chem* 2016;397:709-24.
 97. Cozzo AJ, Fuller AM, Makowski L. Contribution of adipose tissue to development of cancer. *Compr Physiol* 2017;8:237-82.
 98. Petan T, Jarc E, Jusović M. Lipid droplets in cancer: guardians of fat in a stressful world. *Molecules* 2018;23:E1941.
 99. Jarc E, Petan T. Lipid droplets and the management of cellular stress. *Yale J Biol Med* 2019;92:435-52.
 100. Liao Y, Tham DKL, Liang FX, Chang J, Wei Y, Sudhir PR, et al. Mitochondrial lipid droplet formation as a detoxification mechanism to sequester and degrade excessive urothelial membranes. *Mol Biol Cell* 2019;30:2969-84.
 101. Wang CW. Lipid droplets, lipophagy, and beyond. *Biochim Biophys Acta* 2016;1861(8 Pt B):793-805.
 102. Assumpção JAF, Magalhães KG, Corrêa JR. The role of ppar γ and autophagy in ros production, lipid droplets biogenesis and its involvement with colorectal cancer cells modulation. *Cancer Cell Int* 2017;17:82.
 103. Avena P, Anselmo W, Whitaker-Menezes D, Wang C, Pestell RG, Lamb RS, et al. Compartment-specific activation of PPAR γ governs breast cancer tumor growth, via metabolic reprogramming and symbiosis. *Cell Cycle* 2013;12:1360-70.
 104. Wasternack C, Feussner I. The oxylipin pathways: biochemistry and function. *Annu Rev Plant Biol* 2018;69:363-86.

105. Holy P, Kloudova A, Soucek P. Importance of genetic background of oxysterol signaling in cancer. *Biochimie* 2018;153:109-38.
106. Kloudova A, Guengerich FP, Soucek P. The role of oxysterols in human cancer. *Trends Endocrinol Metab* 2017;28:485-96.
107. Mutemberezi V, Guillemot-Legris O, Muccioli GG. Oxysterols: from cholesterol metabolites to key mediators. *Prog Lipid Res* 2016;64:152-69.