

Contribution of oxygen extraction fraction to maximal oxygen uptake in healthy young men

Øyvind Skattebo¹  | Jose A. L. Calbet^{1,2}  | Bjarne Rud¹  | Carlo Capelli^{1,3}  | Jostein Hallén¹ 

¹Department of Physical Performance, Norwegian School of Sport Sciences, Oslo, Norway

²Department of Physical Education and Research Institute of Biomedical and Health Sciences (IUIBS), University of Las Palmas de Gran Canaria, Gran Canaria, Spain

³Department of Neurosciences, Biomedicine and Movement Sciences, University of Verona, Verona, Italy

Correspondence

Øyvind Skattebo, Department of Physical Performance, Norwegian School of Sport Sciences, Post box 4014 Ullevål Stadion, 0806 Oslo, Norway.
Email: oyvind.skattebo@nih.no

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Abstract

We analysed the importance of systemic and peripheral arteriovenous O₂ difference (a- \bar{v} O₂ difference and a-v_fO₂ difference, respectively) and O₂ extraction fraction for maximal oxygen uptake ($\dot{V}O_{2\max}$). Fick law of diffusion and the Piiper and Scheid model were applied to investigate whether diffusion versus perfusion limitations vary with $\dot{V}O_{2\max}$. Articles ($n = 17$) publishing individual data ($n = 154$) on $\dot{V}O_{2\max}$, maximal cardiac output (\dot{Q}_{\max} ; indicator-dilution or the Fick method), a- \bar{v} O₂ difference (catheters or the Fick equation) and systemic O₂ extraction fraction were identified. For the peripheral responses, group-mean data (articles: $n = 27$; subjects: $n = 234$) on leg blood flow (LBF; thermodilution), a-v_fO₂ difference and O₂ extraction fraction (arterial and femoral venous catheters) were obtained. \dot{Q}_{\max} and two-LBF increased linearly by 4.9-6.0 L · min⁻¹ per 1 L · min⁻¹ increase in $\dot{V}O_{2\max}$ ($R^2 = .73$ and $R^2 = .67$, respectively; both $P < .001$). The a- \bar{v} O₂ difference increased from 118-168 mL · L⁻¹ from a $\dot{V}O_{2\max}$ of 2-4.5 L · min⁻¹ followed by a reduction (second-order polynomial: $R^2 = .27$). After accounting for a hypoxemia-induced decrease in arterial O₂ content with increasing $\dot{V}O_{2\max}$ ($R^2 = .17$; $P < .001$), systemic O₂ extraction fraction increased up to ~90% ($\dot{V}O_{2\max}$: 4.5 L · min⁻¹) with no further change (exponential decay model: $R^2 = .42$). Likewise, leg O₂ extraction fraction increased with $\dot{V}O_{2\max}$ to approach a maximal value of ~90-95% ($R^2 = .83$). Muscle O₂ diffusing capacity and the equilibration index Y increased linearly with $\dot{V}O_{2\max}$ ($R^2 = .77$ and $R^2 = .31$, respectively; both $P < .01$), reflecting decreasing O₂ diffusional limitations and accentuating O₂ delivery limitations. In conclusion, although O₂ delivery is the main limiting factor to $\dot{V}O_{2\max}$, enhanced O₂ extraction fraction ($\geq 90\%$) contributes to the remarkably high $\dot{V}O_{2\max}$ in endurance-trained individuals.

Abbreviations: [Hb], haemoglobin concentration; \dot{Q}_{\max} , maximal cardiac output; $\bar{v}O_2$ extraction, systemic oxygen extraction fraction; $\dot{V}O_2$, oxygen uptake; $\dot{V}O_{2\max}$, pulmonary maximal oxygen uptake; a- $\bar{v}O_2$ difference, arterial to mixed venous oxygen difference; $\bar{C}vO_2$, mixed venous oxygen content; a-v_fO₂ difference, arterial to femoral venous oxygen difference; CaO₂, arterial oxygen content; Cv_fO₂, femoral venous oxygen content; CVP, central venous pressure; D_MO₂, muscle O₂ diffusing capacity; LBF, leg blood flow; Leg $\dot{V}O_{2\max}$, maximal oxygen uptake of the leg; MAP, mean arterial blood pressure; MTT, erythrocyte capillary mean transit time; O₂ extraction, peripheral (leg) oxygen extraction fraction; OXPHOS, maximal mitochondrial respiratory capacity; P₅₀O₂, partial pressure of O₂ at 50% SO₂; PO₂, partial pressure of oxygen; SO₂, oxygen saturation of haemoglobin.

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KEYWORDS

arteriovenous oxygen difference, cardiac output, exercise, leg blood flow, limiting factors, maximal oxygen uptake, oxygen diffusion, stroke volume

1 | INTRODUCTION

Under resting conditions in humans, the $\dot{V}O_2$ is 3–5 mL · kg⁻¹ · min⁻¹, and only a small fraction is consumed within the skeletal muscles.¹ However, during incremental exercise, the pulmonary $\dot{V}O_2$ increases gradually and can reach a maximum ($\dot{V}O_{2max}$) of ~90 mL · kg⁻¹ · min⁻¹ depending on gender, age, body weight, genetics, training status and health.^{1–3}

According to the Fick equation, $\dot{V}O_{2max}$ is determined by the product of the maximal cardiac output (\dot{Q}_{max}) and the arterial to mixed venous O_2 difference ($a-\bar{v}O_2$ difference). \dot{Q}_{max} multiplied by the arterial O_2 content (CaO_2) sets the upper limit of systemic O_2 delivery, which is the principal limitation to $\dot{V}O_{2max}$ during exercise recruiting a large muscle mass, at sea level.^{4–6} Despite extensive research since the 1950s on the factors limiting $\dot{V}O_{2max}$, it is still debated whether peripheral O_2 extraction capacity contributes to limiting $\dot{V}O_{2max}$.^{7,8} Several original studies^{4,5,9–12} and review articles^{6,13–15} have addressed this topic in recent decades, yet no study has aimed to statistically analyse all the existing data on the association between $\dot{V}O_{2max}$ and its limiting factors. This kind of analysis is warranted, as the original studies often used homogenous groups with a small number of subjects (<10) since they applied costly and invasive techniques involving catheterizations to determine \dot{Q}_{max} (indicator-dilution techniques or the direct Fick method), regional blood flows (thermodilution or indicator-dilution techniques) and O_2 extraction fraction (calculated by the Fick equation or directly measured through arterial and venous catheters). Consequently, the statistical power is often too low to detect small but meaningful differences between subjects, groups with different training status and before and after training, thus precluding a definite conclusion.

It is documented that the $a-\bar{v}O_2$ difference at $\dot{V}O_{2max}$ is only slightly different between untrained and endurance-trained individuals,^{16,17} suggesting that peripheral adaptations to endurance training have only a minor impact on $\dot{V}O_{2max}$. However, the $C\bar{v}O_2$ difference is determined not only by the peripheries' ability to extract O_2 , reflected in the mixed venous O_2 content ($\bar{v}O_2$) but also by the CaO_2 , which sets the upper limit for the $a-\bar{v}O_2$ difference during maximal exercise. The CaO_2 is set by the haemoglobin concentration ([Hb]) and the O_2 saturation of Hb (SO_2), which may change with training and is acutely modified during exercise. For instance, endurance training causes plasma volume expansion¹⁸ that can lead to haemodilution and a lower O_2 carrying capacity of the blood.¹⁶ A high \dot{Q}_{max} shortens the time

for alveolar/capillary gas equilibration at the lung causing exercise-induced arterial hypoxemia that further reduces the CaO_2 .^{19,20} Therefore, it may be that the $a-\bar{v}O_2$ difference does not increase substantially after endurance training because of a concurrent training-induced lowering of CaO_2 , whereas the systemic O_2 extraction fraction (\bar{O}_2 extraction: $a-\bar{v}O_2$ difference/ CaO_2) may improve.

Another aspect of this discussion is whether the measurement techniques are sensitive enough to detect meaningful changes in the $a-\bar{v}O_2$ difference. Most studies have not measured $a-\bar{v}O_2$ difference directly but calculated it using the Fick equation ($\dot{V}O_{2max}/\dot{Q}_{max}$).^{16,17,21–24} The reason why so few studies have measured $a-\bar{v}O_2$ difference directly during maximal exercise is because of the need for right heart catheterization. Therefore, studies measuring the arterial to femoral venous O_2 difference ($a-v_fO_2$ difference) and leg O_2 extraction fraction directly using peripheral catheters may be more sensitive in evaluating whether the O_2 extraction capacity changes with endurance training.

It is important to note that the factors limiting $\dot{V}O_{2max}$ may change over the course of training. For instance, the maximal mitochondrial respiratory capacity (OXPHOS) measured in permeabilized muscle fibres *ex vivo* and $\dot{V}O_{2max}$ is associated in untrained, but not in trained individuals.²⁵ These and other data²⁶ suggest that peripheral factors contribute to limit $\dot{V}O_{2max}$ in the untrained state, but their influence may diminish with increased $\dot{V}O_{2max}$ and training status.

In the present study, we critically reviewed and statistically analysed the previously published data on the association between $\dot{V}O_{2max}$ and O_2 extraction fraction, in men, by focusing on catheterization studies. Two approaches were used: Part 1) articles containing individual data on pulmonary $\dot{V}O_{2max}$, \dot{Q}_{max} (indicator-dilution techniques or the Fick method), $a-\bar{v}O_2$ difference (mostly calculated) and \bar{O}_2 extraction fraction measured during whole-body maximal exercise (running, cycling) were included; Part 2) to investigate the relationship between limb $\dot{V}O_2$ and peripheral O_2 extraction fraction, mean data from studies reporting leg blood flow (LBF), $a-v_fO_2$ difference and leg O_2 extraction fraction (catheters) measured during whole-body maximal exercise (running, cycling, cross-country skiing) were included. To investigate whether the limiting factors vary with $\dot{V}O_{2max}$, we employed the Fick law of diffusion to calculate the muscle O_2 diffusing capacity (D_MO_2) and subsequently used the Piiper and Scheid model to calculate the relative roles of perfusion versus diffusion limitations to $\dot{V}O_{2max}$.²⁷ Finally, we discuss the potential mechanisms behind the elevated O_2 extraction fraction observed after endurance training.

2 | ANALYSIS OF EXISTING DATA

The strategy to use individual and mean data to investigate the systemic and peripheral responses, respectively, was chosen since a large amount of individual data has been published on systemic responses, whereas we were unable to identify other than mean values in studies investigating peripheral haemodynamics and O_2 extraction fraction. The data were identified through searches conducted in the PubMed database using several combinations of the following search terms: circulation, circulatory, hemodynamic(s), cardiac output, leg blood flow, arteriovenous oxygen difference, oxygen extraction and exercise. Cross-reference checks were also conducted, in addition to separate searches on authors with articles already included in the database. Only exercise modes engaging a large muscle mass that could elicit $\dot{V}O_{2max}$ were included (cycling, running and cross-country skiing using the diagonal technique). Data from cross-sectional studies or before and after training interventions that were collected in normoxia on young (<40 years old) and healthy individuals were included. Data collected in hypoxia, after acclimatization to altitude, in altitude natives, in hyperthermia, with atrial pacing,

after bed rest and after blood volume manipulations were excluded. The control condition was used when the above forms of manipulations of the cardiovascular system were conducted. Only catheterization studies that used invasive methods to measure \dot{Q}_{max} (indicator-dilution techniques or the Fick method) and LBF (bolus or continuous infusion thermodilution and indicator-dilution techniques) were included. Only individual data from men are used (Part 1). In Part 2, studies that had a sample with a majority of men were used ($\geq 50\%$). When several papers reported data from the same data collection, only one of the articles was included. If an article used some of the same subjects as previously reported, but with supplementation with new subjects, the data were included. The included articles are presented in Tables 1 and 2 for Parts 1 and 2, respectively.

2.1 | Calculations

When the data were published in graphs and not in tables or text, ImageJ (v1.50b; National Institutes of Health, USA) was used for data extraction. If not all variables were reported

TABLE 1 Articles reporting individual values of maximal oxygen uptake ($\dot{V}O_{2max}$), maximal cardiac output (\dot{Q}_{max}) and arterial to mixed venous O_2 difference (a- $\bar{v}O_2$ difference). In studies reporting arterial O_2 content, the systemic O_2 extraction fraction (O_2 extraction) was calculated

Article	n	Exercise	Age	Method used to measure:			Reported or can be calculated		
				\dot{Q}_{max}	$\dot{V}O_{2max}$	a- $\bar{v}O_2$ difference	$\bar{v}O_2$ extraction	MAP	
Blomqvist et al ¹¹⁶	4	Cycling	23-33	ID	DB	Calculated using Fick equation ↑	–	Yes	
Eklblom and Hermansen ¹⁶	14	Run	22-34	ID	DB		Yes	Yes	
Eklblom et al ¹⁷	8	Cycling	19-27	ID	DB		Yes	Yes	
Eklblom ¹¹²	7	Cycling	22-26	ID	DB		Yes	Yes	
Epstein et al ¹¹⁷	2	Run	21	ID	Custom	Measured ↓	–	Yes	
Epstein et al ¹¹⁸	4	Run	18-30	Fick	Custom		–	Yes	
Gleser ²²	6	Cycling	20-23	ID	DB		Calculated using Fick equation	Yes	Yes
Hermansen et al ²¹	13	Cycling/Run	19-34	ID	DB		Yes	Yes	
Mitchell et al ²³	6	Run	–	ID	DB		–	–	
Robinson et al ¹¹⁹	5	Run	19-31	ID	Custom		–	Yes	
Saltin ¹¹⁰	4	Cycling	23-26	ID	DB		–	–	
Saltin and Stenberg ¹⁰⁹	4	Cycling	23-25	ID	DB		–	Yes	
Saltin et al ⁸⁵	5	Cycling	19-21	ID	DB		Yes	Yes	
Saltin et al ¹¹¹	4	Cycling	20-21	ID	DB		–	–	
Stenberg et al ¹¹⁴	6	Cycling	20-36	ID	DB		Yes	Yes	
Stenberg et al ¹⁰⁸	5	Cycling	20-39	ID	DB		–	Yes	
Åstrand et al ²⁴	12	Cycling	21-30	ID	DB		Yes	Yes	

Abbreviations: DB, Douglas bag technique; ID, indicator-dilution method using indocyanine green or Evans blue dye (only used in Mitchell et al²³); MAP, mean arterial pressure; n, number of subjects meeting the inclusion criteria. Note that some subjects were investigated on more than one occasion (before/after training, running/cycling).

TABLE 2 Articles reporting mean values of leg oxygen uptake (leg $\dot{V}O_{2\max}$), leg blood flow (LBF) and arterial to femoral venous O_2 difference ($a-v_fO_2$ difference) during maximal exercise (cycling and cross-country skiing using the diagonal technique)

Article	n	Age (\bar{x})	Method used to measure:			Reported or can be calculated
			Pulmonary $\dot{V}O_2$	LBF	a- v_fO_2 difference	O_2 extraction
Bender et al ¹⁰²	7♂	22	Custom	TD-B	Measured via arterial and femoral venous blood sampling ↓	Yes
Calbet et al ⁵⁷	4♂3♀	24	Med. Graph. CPX	TD-C		Yes
Calbet et al ^{29,32}	3♂	24	Amis 2001	TD-C		Yes
Calbet et al ⁵²	10♂	24	Quark b2	TD-C		Yes
Calbet et al ⁵	9♂	33	Quark b2	TD-C		Yes
Calbet et al ³¹	9♂	31	Quark b2	TD-C		Yes
Calbet et al ⁵⁶	11♂	22	Vmax 29	TD-C		Yes
Cardinale et al ¹⁰	4♂3♀	33	Oxycon Pro	TD-C		Yes
Cardus et al ¹⁰⁴	13♂5♀	23	Custom	TD-C		Yes
Gonzalez-Alonso et al ¹¹	8♂	24	OCM-2	TD-C		Yes
Harms et al ¹⁰⁵	7♂	29	Custom	TD-C	Yes	
Klausen et al ³⁰	6♂	23	Douglas bag tech.	ID-B	–	
Knight et al ¹⁰³	7♂	29	Custom	TD-C	Yes	
Knight et al ⁹⁷	12♂	29	Custom	TD-C	Yes	
Lundby et al ⁵³	8♂	26	Quark b2	TD-C	Yes	
Lundby et al ⁵⁴	8♂	27	Quark b2	TD-C	Yes	
Lundby et al ^{107,113}	6♂	26	Custom	TD-C	Yes	
Mortensen et al ⁹	13♂	28	Quark b2	TD-C	Yes	
Mortensen et al ⁴	10♂	27	Quark b2	TD-C	Yes	
Munch et al ⁵⁵	10♂	27	Quark CPET	TD-C	Yes	
Poole et al ¹¹⁵	6♂	26	Custom	TD-C	Yes	
Roca et al ²⁸	6♂	24	Custom	–	Yes	
Roca et al ¹²	8♂4♀	22	Custom	TD-C	Yes	
Proctor et al ¹²⁰	11♂	21	TrueMax 2400	TD-C	Yes	
Rud et al ⁴⁵	4♂4♀	23	Douglas bag tech.	TD-C	Yes	
Trangmar et al ⁷⁶	9♂	26	Not reported	TD-C	Yes	
van Hall et al ¹⁰⁶	5♂1♀	26	Med. Graph. CPX	TD-C	Yes	

Abbreviations: ID-B, bolus indicator-dilution method (I-labelled human albumin); n, number of subjects; TD-B, bolus-infusion thermodilution method; TD-C, continuous-infusion TD.

in the articles, the reported data were used to derive the missing values via the following formulas or combination of formulas if possible:

$$\text{Pulmonary } \dot{V}O_{2\max} = \dot{Q}_{\max} \times a-\bar{v}O_2 \text{ difference} \quad (1)$$

$$\text{Leg } \dot{V}O_{2\max} = \text{LBF} \times a-v_fO_2 \text{ difference} \quad (2)$$

$$\text{Stroke volume} = \dot{Q}_{\max} / \text{heart rate} \quad (3)$$

$$\text{Blood } O_2 \text{ content (e.g., } CaO_2) = 1.39 \times [\text{Hb}] \times SO_2 + 0.003 \times PO_2 \quad (4)$$

$$\text{Leg } O_2 \text{ delivery} = \text{LBF} \times CaO_2 \quad (5)$$

$$\bar{O}_2 \text{ extraction} = a-\bar{v}O_2 \text{ difference} / CaO_2 \quad (6)$$

$$O_2 \text{ extraction} = a-v_fO_2 \text{ difference} / CaO_2 \quad (7)$$

$$\text{Systemic vascular conductance} = \dot{Q}_{\max} / (\text{MAP} - \text{CVP}) \quad (8)$$

If no arterial partial pressure of O_2 (PO_2) was reported, 100 mmHg was assumed for the calculation of CaO_2 (ie, 3 mL O_2 freely dissolved in blood plasma per 1 L of blood). Central venous pressure (CVP) at $\dot{V}O_{2\max}$ was taken as 5 mmHg⁴ when calculating systemic vascular conductance.

$D_M O_2$ and mean capillary PO_2 were calculated as previously described,^{28,29} using the measured arterial and femoral venous PO_2 . $D_M O_2$ [$D_M O_2 = \dot{V}O_{2}/(\text{mean capillary } PO_2 - \text{mitochondrial } PO_2)$]; ie, the O_2 conductance from the capillary to the mitochondria] is recognized as a compound variable integrating several steps in the O_2 cascade, including the dissociation of O_2 from Hb, and diffusion through the erythrocyte membrane, plasma, capillary wall, interstitial space, sarcolemma, cytoplasm (myoglobin facilitated or by diffusion) and into the mitochondria for utilization by the cytochromes. The equilibration index Y , which quantitatively describes perfusion versus diffusion limitations to $\dot{V}O_2$, was calculated according to Piiper and Scheid.²⁷

2.2 | Statistical analyses

Data are presented as mean \pm standard deviation, if not otherwise stated. Regression was analysed using simple linear regression, second-order polynomials and exponential decay models ($y = a \cdot e^{-K \cdot X} + \text{plateau}$), all using least squares as the fitting method. Regression lines/curves are presented with 95% confidence bands representing the likely location of the true curve. The alpha-level was set to $\leq .05$ and values between $> .05$ and $\leq .10$ were considered to indicate trends. GraphPad Prism (v. 8.0.1; GraphPad Software, CA, USA) and Microsoft Office Excel 2013 (Microsoft Corporation, WA, USA) were used for statistical analysis.

2.3 | Part 1: Systemic responses during maximal exercise (individual data)

\dot{Q}_{max} increased by $4.9 \text{ L} \cdot \text{min}^{-1}$ for each $\text{L} \cdot \text{min}^{-1}$ increase in $\dot{V}O_{2\text{max}}$ (Figure 1A; $P < .001$), explained by a linear increase in stroke volume (Figure 1B; $P < .001$).

The calculated a- $\bar{v}O_2$ difference ($\dot{V}O_{2\text{max}}/\dot{Q}_{\text{max}}$) showed an inverse J-shaped curve, reaching the highest level between $4.5\text{--}5.0 \text{ L} \cdot \text{min}^{-1}$ before declining at higher $\dot{V}O_{2\text{max}}$ (Figure 1C). After accounting for the decrease in CaO_2 with increasing $\dot{V}O_{2\text{max}}$ (Figure 1E; $P < .001$), the calculated \bar{O}_2 extraction fraction increased up to a $\dot{V}O_{2\text{max}}$ of $\sim 4.5\text{--}5.0 \text{ L} \cdot \text{min}^{-1}$ and then approached a maximal value at $\sim 90\%$ (Figure 1D) when restricting the exponential decay model to plausible physiological limits ($\dot{V}O_{2\text{max}}$: $6\text{--}7 \text{ L} \cdot \text{min}^{-1}$). The linear decrease in CaO_2 was explained by arterial hypoxemia (decreased arterial SO_2 ; Figure 1F; $P < .001$) and a non-significant negative relationship between [Hb] and $\dot{V}O_{2\text{max}}$ (Figure 1G; $P = .232$). The calculated $C\bar{v}O_2$ gradually decayed and approached a minimum at $\sim 10\text{--}15 \text{ mL} \cdot \text{L}^{-1}$ in the subjects with the highest $\dot{V}O_{2\text{max}}$ (Figure 1H).

Systemic vascular conductance was strongly positively correlated with $\dot{V}O_{2\text{max}}$ (Figure 2B; $P < .001$). There were

no significant associations between mean arterial pressure (MAP) and $\dot{V}O_{2\text{max}}$ (Figure 2A; $P = .289$) or with \dot{Q}_{max} ($y = -0.2x + 125$; $R^2 = .004$; $n = 119$; $P = .475$).

When controlling the regression between the individual data of $\dot{V}O_{2\text{max}}$ and the calculated \bar{O}_2 extraction fraction with mean values from studies measuring \bar{O}_2 extraction fraction directly using the Fick method (right heart catheterization), or indirectly using the Fick equation (\dot{Q}_{max} : indicator-dilution or transpulmonary thermodilution), most values fell close to the regression curve (Figure 3).

2.4 | Part 2: Peripheral responses during maximal exercise (mean data)

LBF and two-LBF rose by 4.6 and $5.7 \text{ L} \cdot \text{min}^{-1}$ for each $\text{L} \cdot \text{min}^{-1}$ increase in leg and pulmonary $\dot{V}O_{2\text{max}}$ respectively (Figure 4A,D; both $P < .001$). Leg and pulmonary $\dot{V}O_{2\text{max}}$ displayed a linear relationship ($y = 1.27x - 2.01$; $R^2 = .85$; $n = 28$; $P < .001$). The directly measured leg a- $v_f O_2$ difference and leg O_2 extraction fraction were best explained by exponential decay models and increased gradually with the increase in leg and pulmonary $\dot{V}O_{2\text{max}}$ to approach a maximum at $\sim 180\text{--}190 \text{ mL} \cdot \text{L}^{-1}$ and $\sim 90\text{--}95\%$ respectively (Figure 4B,C,E,F). These relationships were equally strong when $\dot{V}O_{2\text{max}}$ was standardized to body weight (Supporting material Figure 1). Note that leg a- $v_f O_2$ difference was not lower for the subjects with the highest $\dot{V}O_{2\text{max}}$, as observed for the systemic a- $\bar{v}O_2$ difference (Figure 1C), possibly since only one subject group exceeded a $\dot{V}O_{2\text{max}}$ of $4.7 \text{ L} \cdot \text{min}^{-1}$, where this occurred for the systemic responses (see Figure 1C). In connection, no association was evident between pulmonary $\dot{V}O_{2\text{max}}$ and CaO_2 for these data ($y = 1.07 + 195$; $R^2 < .01$; $n = 30$; $P = .701$).

Like the systemic responses, the measured femoral venous O_2 content ($Cv_f O_2$) decreased gradually with increasing pulmonary $\dot{V}O_{2\text{max}}$ until reaching a minimum of $\sim 10 \text{ mL} \cdot \text{L}^{-1}$ (Figure 5A). Likewise, the femoral venous SO_2 and PO_2 decreased gradually to approach $\sim 5\%$ and $\sim 10 \text{ mmHg}$ at the highest $\dot{V}O_{2\text{max}}$ respectively (Figure 5B,C).

$D_M O_2$ was positively correlated with leg $\dot{V}O_{2\text{max}}$ ($y = 27x - 6$; $R^2 = .92$; $n = 21$; $P < .001$), pulmonary $\dot{V}O_{2\text{max}}$ (Figure 6A; $P < .001$) and leg O_2 extraction fraction ($y = 1.7x - 110$; $R^2 = .80$; $n = 21$; $P < .001$). Interestingly, the equilibration index Y , which quantitatively describes diffusion versus perfusion limitations to muscle $\dot{V}O_2$ (where $Y < 0.1$ indicates pure diffusion limitation, $0.1 < Y < 3$ indicates mixed perfusion-diffusion limitation and $Y > 3$ indicates pure perfusion limitation),²⁷ was well above 1.0 for all subject groups (Figure 6B) and increased progressively with leg $\dot{V}O_{2\text{max}}$ ($y = 0.28x + 1.40$; $R^2 = .37$; $n = 21$; $P = .003$), pulmonary $\dot{V}O_{2\text{max}}$ (Figure 6B; $P = .008$) and leg O_2 extraction fraction ($y = 0.023x - 0.129$; $R^2 = .53$; $n = 21$; $P < .001$). The equilibration index Y was also correlated with pulmonary $\dot{V}O_{2\text{max}}$

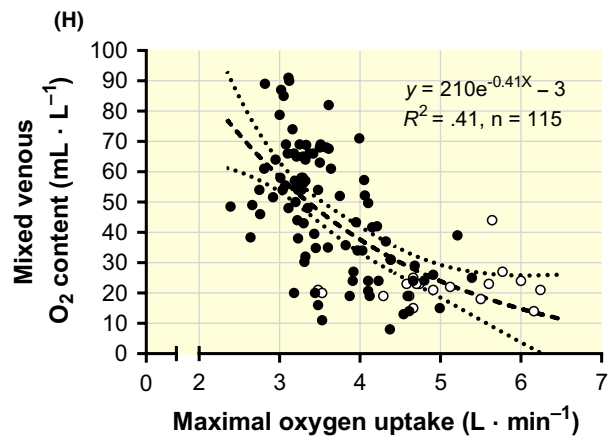
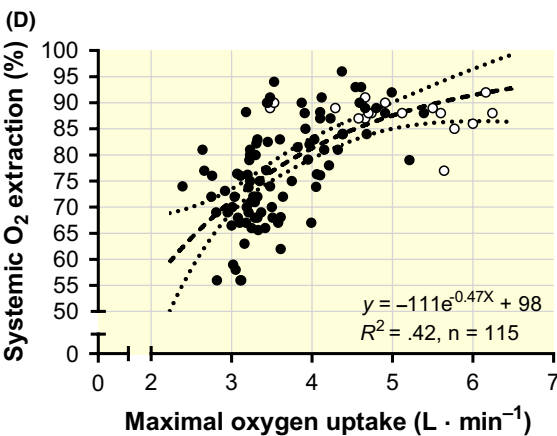
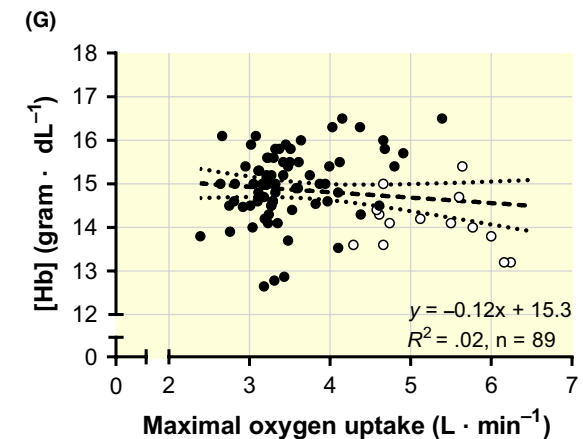
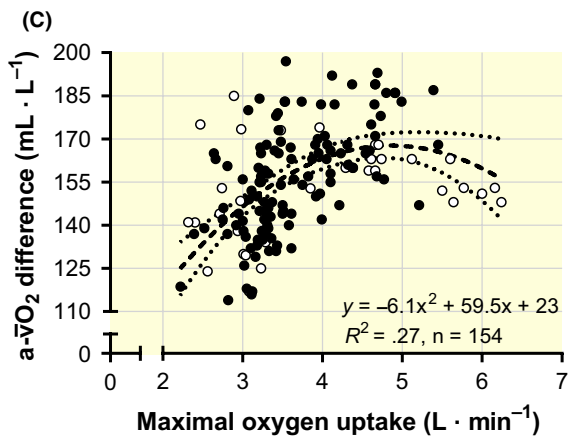
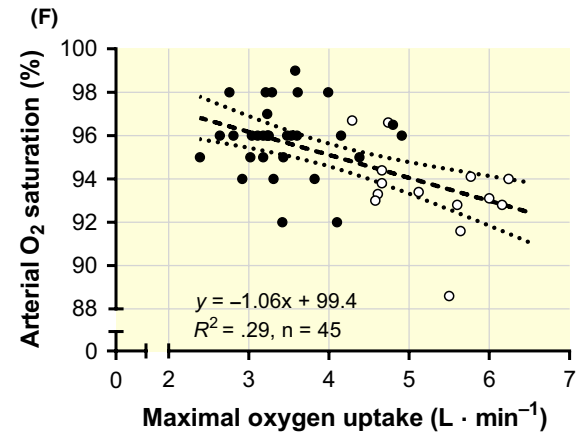
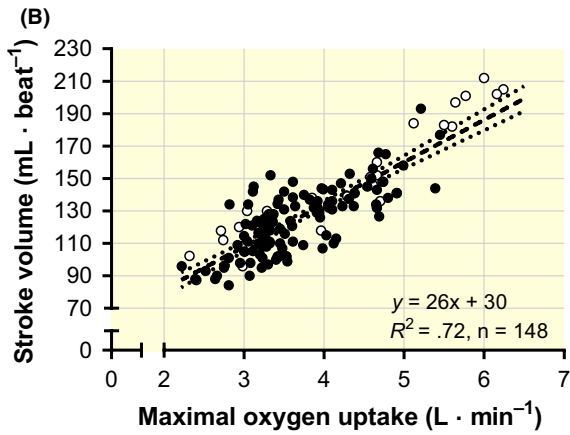
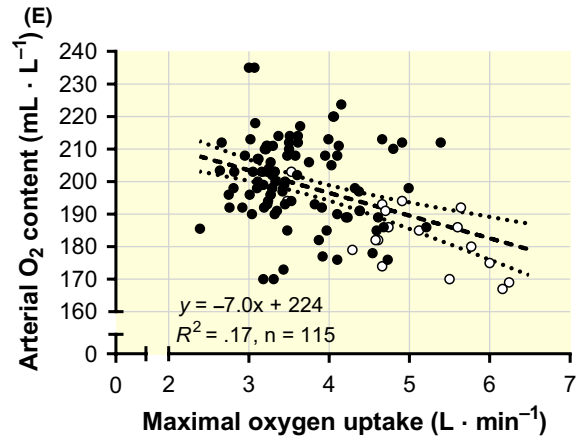
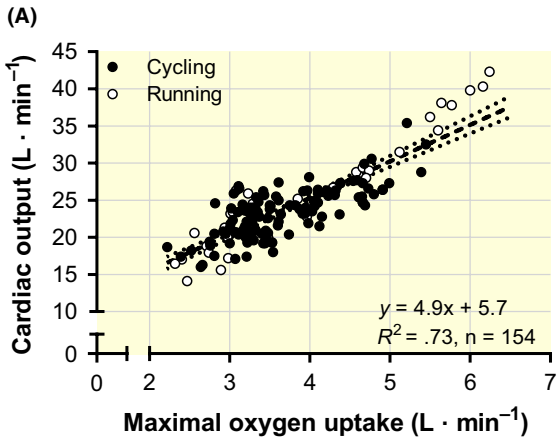


FIGURE 1 The relationship between individual values (from studies reported in Table 1) of pulmonary maximal oxygen uptake and cardiac output (A), stroke volume (B), arterial to mixed venous oxygen difference ($a-\bar{v}O_2$ difference; C), systemic oxygen extraction fraction (D), arterial oxygen content (E), arterial oxygen saturation (F), haemoglobin concentration ([Hb]; G) and the calculated mixed venous oxygen content (H). All data were obtained during maximal exercise. Inserted in each graph are the formulas for the regression equations along with the goodness of fit (R^2) and the number of data pairs (n)

standardized to body weight ($R^2 = .38$; $P = .003$; Supporting material Figure 2). Therefore, the leg muscles were more perfusion than diffusion limited, even for subjects with the lowest $\dot{V}O_{2\max}$, and were progressively more perfusion/ O_2 delivery limited with a gradually higher $\dot{V}O_{2\max}$. This can also be illustrated by applying the Piiper and Scheid model to calculate the fractional extent to which $\dot{V}O_{2\max}$ is expected to change if $D_M O_2$ or LBF are modified²⁷; Figure 6C shows that an individual's $\dot{V}O_{2\max}$ is less sensitive to any change in $D_M O_2$ if the $\dot{V}O_{2\max}$ is already high, which is caused by the little remaining O_2 available for extraction in the femoral venous (ie, end-capillary) blood. For instance, according to this

theoretical model and using the relationship in Figure 6C; if a subject with a $\dot{V}O_{2\max}$ of $5 \text{ L}\cdot\text{min}^{-1}$ changed his $D_M O_2$ by 20%, he would only change his $\dot{V}O_{2\max}$ by $\sim 6\%$ ($20\% \times 0.3$). Conversely, the same subject would increase $\dot{V}O_{2\max}$ by $\sim 14\%$ after a 20% increase in LBF ($20\% \times 0.7$).

3 | SUMMARY OF FINDINGS

To our knowledge, the present investigation is the first to critically review the existing research on the association between $\dot{V}O_{2\max}$ and systemic and peripheral O_2 extraction fractions in healthy young men. Our findings are as follows:

1. Pulmonary and leg $\dot{V}O_{2\max}$ were best explained by \dot{Q}_{\max} and LBF, respectively, agreeing with most previous studies where these variables have been directly manipulated.
2. The systemic \bar{O}_2 extraction fraction increased with $\dot{V}O_{2\max}$ until approximately $4.5\text{--}5.0 \text{ L}\cdot\text{min}^{-1}$. Above this value, the \bar{O}_2 extraction fraction was typically around $\sim 90\%$.
3. The measured leg O_2 extraction fraction increased with leg and pulmonary $\dot{V}O_{2\max}$ to approach a maximal value at $\sim 90\text{--}95\%$, strengthening the findings from the calculated systemic \bar{O}_2 extraction fraction. This strongly suggests that O_2 extraction increases after endurance training and contributes to a high $\dot{V}O_{2\max}$.
4. The calculated $C\bar{v}O_2$ and the measured $Cv_f O_2$ indicate a minimum value at ~ 15 and $\sim 10 \text{ mL}\cdot\text{L}^{-1}$, respectively, associated with a femoral venous SO_2 and PO_2 of $\sim 5\%$ and $\sim 10 \text{ mmHg}$ respectively. At this point, further peripheral O_2 extraction may no longer be possible as a result of diffusional limitations and/or because the remaining O_2 represents blood perfusing the least active muscle regions of the leg, connective tissue, bone marrow, adipose tissue and skin, which are characterized by a lower O_2 extraction.
5. The progressive increase in the equilibration index Y with pulmonary and leg $\dot{V}O_{2\max}$ indicates that the muscles become gradually more perfusion/ O_2 delivery limited with increasing $\dot{V}O_{2\max}$.

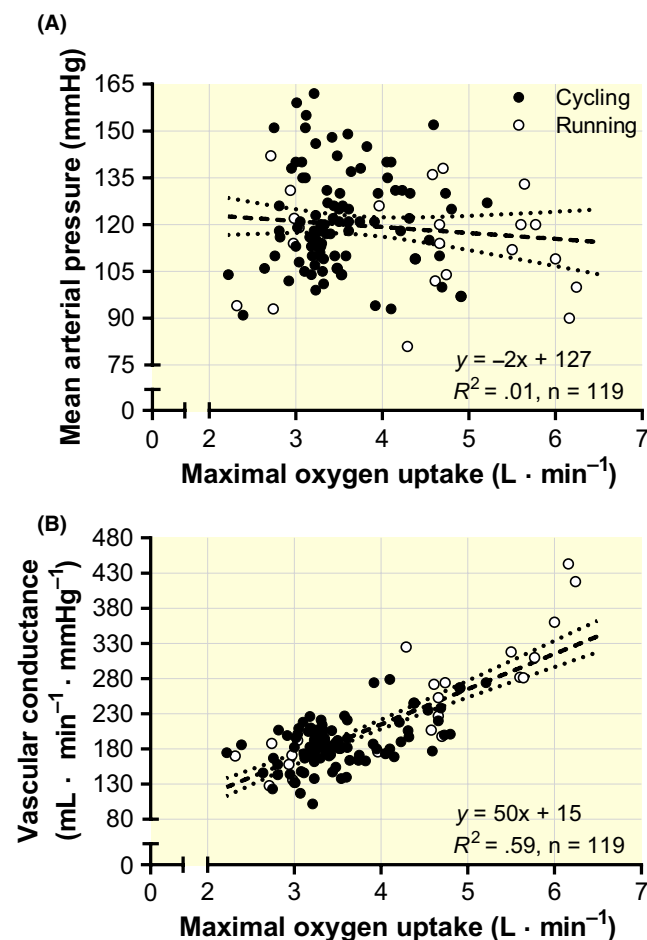


FIGURE 2 The relationship between individual values (from studies reported in Table 1) of pulmonary maximal oxygen uptake and mean arterial pressure (A) and systemic vascular conductance (B). Inserted in each graph are the formulas for the linear regression along with the goodness of fit (R^2) and the number of data pairs (n)

3.1 | Oxygen delivery

To match O_2 delivery to O_2 consumption, \dot{Q}_{\max} and two-LBF increased by $\sim 5\text{--}6 \text{ L}\cdot\text{min}^{-1}$ per $1 \text{ L}\cdot\text{min}^{-1}$ increase in pulmonary $\dot{V}O_{2\max}$. These relationships were strong and complied

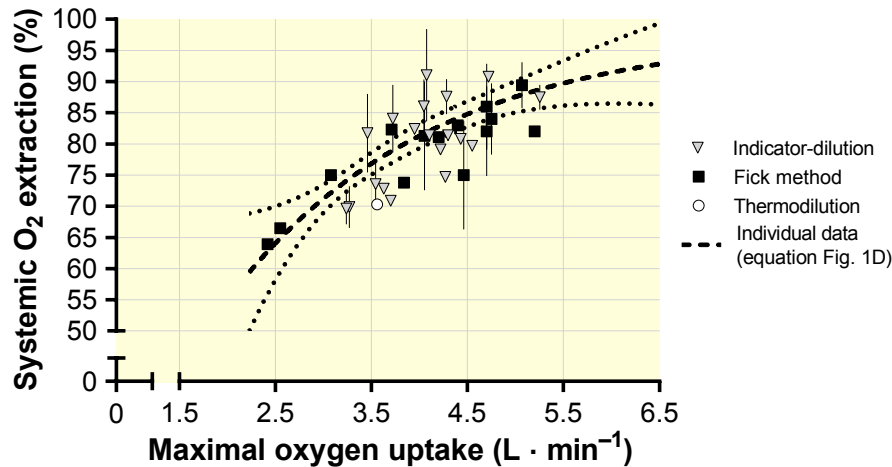


FIGURE 3 Mean values ($\pm 95\%$ confidence limits, where available) of systemic oxygen extraction fraction versus maximal oxygen uptake from studies using the direct (pulmonary artery catheter) or the modified (right atrium catheter) Fick method,^{4,9,28,32,51,55,61,121-127} the indicator dilution method^{5,11,16,17,21,22,24,31,42,52-54,57,85,112,114,128-130} and the transpulmonary thermodilution method.⁵⁶ Broken line is the regression equation obtained from Figure 1D

with previous research and the “classic” view that O_2 delivery is the primary determinant of whole-body $\dot{V}O_{2max}$.^{4,7,11} As maximal heart rate showed no apparent relationship with $\dot{V}O_{2max}$, the high stroke volumes ($>180 \text{ mL} \cdot \text{beat}^{-1}$) explained the large \dot{Q}_{max} in the athletes included in the present analysis ($>35 \text{ L} \cdot \text{min}^{-1}$), in agreement with previous knowledge.^{13,16,30}

Despite increased \dot{Q}_{max} , MAP was unchanged with increasing $\dot{V}O_{2max}$ as a result of increased vascular conductance. Although untrained individuals typically display a rise in MAP from rest to maximal exercise,³¹ well-trained athletes can display an unchanged MAP or even a small reduction owing to profound peripheral vasodilation.³² Consequently, vasodilation of a well-developed peripheral vascular network likely contributed to the extremely high stroke volumes by minimizing afterload in the subjects with the highest $\dot{V}O_{2max}$. To substantiate, endurance training of each leg separately, to evoke extensive peripheral adaptations without stimulating the central circulation substantially, has been shown to decrease MAP and the total peripheral resistance during two-legged maximal exercise that likely contributed to the elevated stroke volume and \dot{Q}_{max} after training.³⁰ The high stroke volumes are probably achieved through the combined effect of a large left ventricular mass,^{33,34} compliant cardiac chambers^{35,36} and an expanded blood volume^{37,38} that facilitates a high end-diastolic volume and preload combined with the relatively low afterload.

3.2 | Oxygen extraction

The calculated systemic $a-\bar{v}O_2$ difference showed a large variability for a given $\dot{V}O_{2max}$ and was, if anything, lower in those subjects displaying the highest $\dot{V}O_{2max}$ ($>5 \text{ L} \cdot \text{min}^{-1}$) compared to those being moderately to well trained ($\dot{V}O_{2max}$:

$4\text{-}5 \text{ L} \cdot \text{min}^{-1}$). This agrees with previous studies showing only a small difference between non-endurance-trained and active individuals^{16,17} and no apparent difference between well-trained individuals and elite athletes.¹⁶ This has led previous investigators to argue that improved O_2 extraction does not contribute or only minimally contributes to the remarkably high $\dot{V}O_{2max}$ observed in elite athletes.^{14,39} However, these papers may not have considered that endurance training causes plasma volume expansion,¹⁸ which often leads to haemodilution and a lower O_2 carrying capacity of the arterial blood.¹⁶ Combined with the below-average haemoconcentration from rest to maximal exercise that occurs in well-trained individuals¹⁶ and the exercise-induced arterial hypoxemia that often accompanies a high \dot{Q}_{max} ,^{19,20} individuals with the highest $\dot{V}O_{2max}$ displayed a substantially lower CaO_2 ($\sim 10\%$) than those with a low $\dot{V}O_{2max}$ ($<180 \text{ mL} \cdot \text{L}^{-1}$ vs $>200 \text{ mL} \cdot \text{L}^{-1}$; Figure 1E). Therefore, the lower CaO_2 may explain why moderately and well-trained individuals can have a similar $a-\bar{v}O_2$ difference, despite differing markedly in $D_M O_2$, mitochondrial mass and capillary density.^{40,41} Actually, parts of this mechanism are demonstrated experimentally since acute plasma volume expansion increases \dot{Q}_{max} but lowers the CaO_2 and, hence, reduces the $a-\bar{v}O_2$ difference during maximal exercise.^{42,43}

Opposite to the $a-\bar{v}O_2$ difference, the systemic \bar{O}_2 extraction fraction—ie, the fraction of O_2 that is taken up with respect to the amount available for utilization ($a-\bar{v}O_2$ difference/ CaO_2)—increased with $\dot{V}O_{2max}$ until reaching $\sim 90\%$. This pattern was confirmed in the leg when measured using catheters, with the O_2 extraction fraction increasing progressively with leg and pulmonary $\dot{V}O_{2max}$ until reaching ~ 90 to 95% . Therefore, the calculated systemic \bar{O}_2 extraction fraction (Fick equation) is supported by direct measurements via arterial and femoral venous blood sampling and strongly

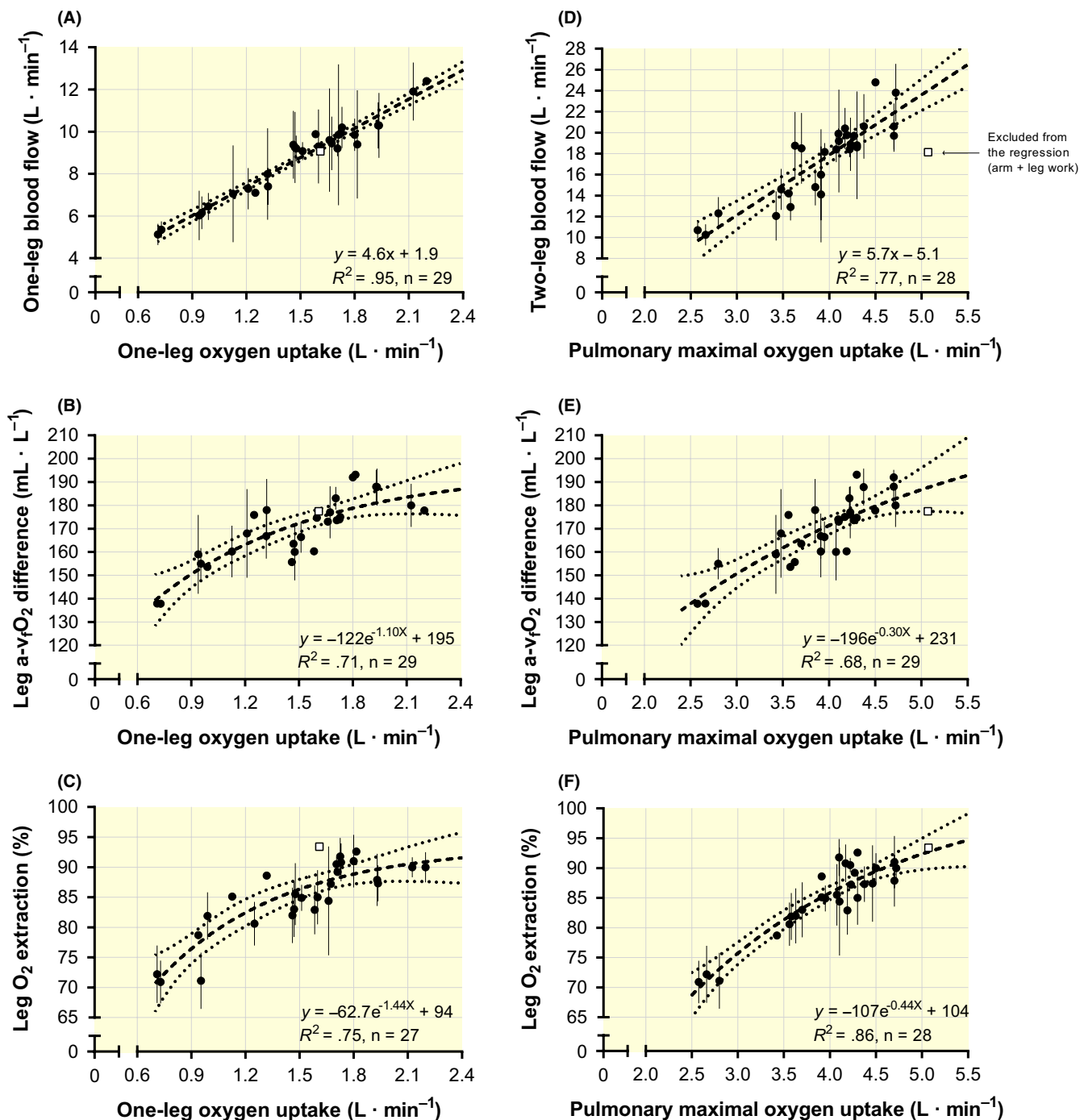


FIGURE 4 The relationship between one-leg or pulmonary maximal oxygen uptake and leg blood flow (Figure 4A,D, respectively), arterial to femoral venous oxygen difference ($a-v_f\text{O}_2$ difference; Figure 4B,E, respectively) and leg oxygen extraction fraction (Figure 4C,F, respectively). Black circles and white squares denote cycling and diagonal cross-country skiing respectively. The skiers are excluded from the regression in Figure 4D owing to the combined leg and arm use for locomotion that distributed $6.6 L \cdot \text{min}^{-1}$ blood flow to the exercising arms (see the discussion). Data are mean values ($\pm 95\%$ confidence limits, where available) from studies reported in Table 2

indicates that the $\bar{\text{O}}_2$ extraction fraction is improved with increasing $\dot{V}\text{O}_{2\text{max}}$ to a certain level.

In most endurance training studies investigating the interplay between central and peripheral adaptations in improving $\dot{V}\text{O}_{2\text{max}}$, \dot{Q}_{max} was measured by non-invasive methods (such as inert-gas rebreathing techniques, impedance cardiography

and bioelectance) and the Fick equation was used to derive the $a-v\bar{\text{O}}_2$ difference (for references, see the meta-analysis by Montero et al⁴⁴). The majority of these studies failed to detect a statistically significant change in the $a-v\bar{\text{O}}_2$ difference. However, this finding does not necessarily mean that $\dot{V}\text{O}_{2\text{max}}$ was exclusively increased by elevated \dot{Q}_{max} for three

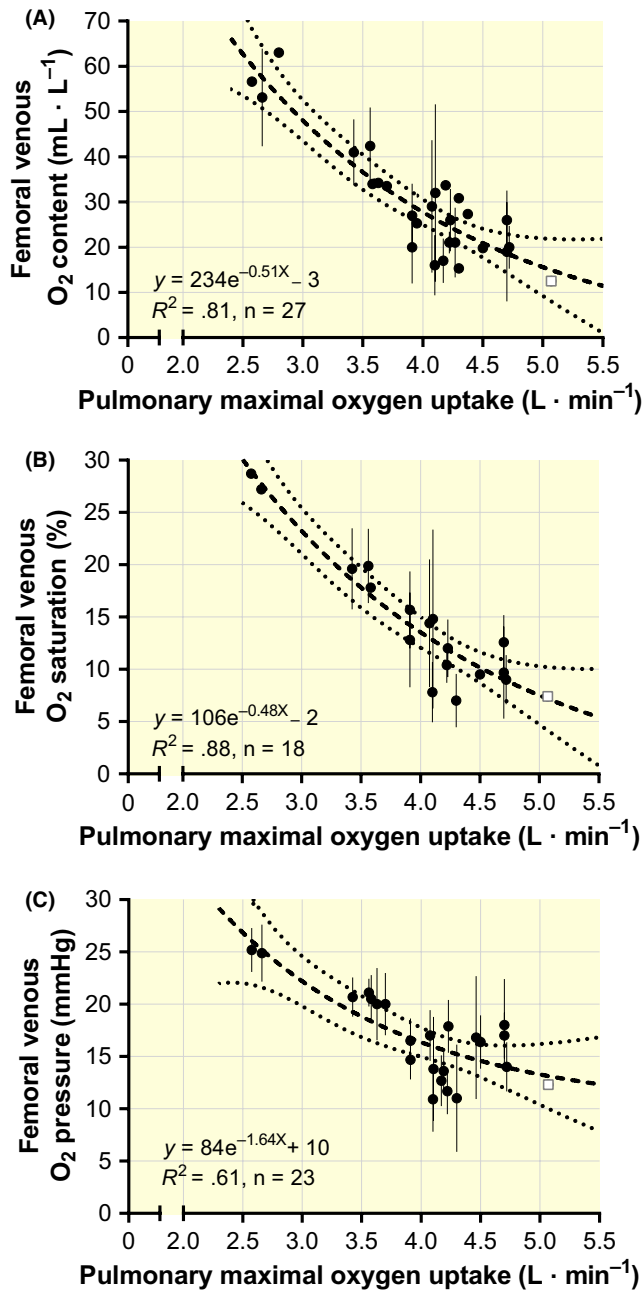


FIGURE 5 The relationship between pulmonary maximal oxygen uptake and the associated femoral venous O₂ content (A), femoral venous O₂ saturation (B) and femoral venous O₂ pressure (C). Black and white symbols denote cycling and diagonal skiing, respectively. Data are mean values ($\pm 95\%$ confidence limits, where available) from studies reported in Table 2

reasons. First, when the $a\text{-}\bar{v}O_2$ difference is calculated by the Fick equation, a large variation is introduced as a result of measurement error in \dot{Q}_{\max} , especially when non-invasive methods are used. Second, because of the above, maybe in combination with a considerable individual variation in peripheral adaptations such as capillarization, it is likely that these studies are underpowered for detecting small changes in the $a\text{-}\bar{v}O_2$ difference. Third, these studies may have failed to

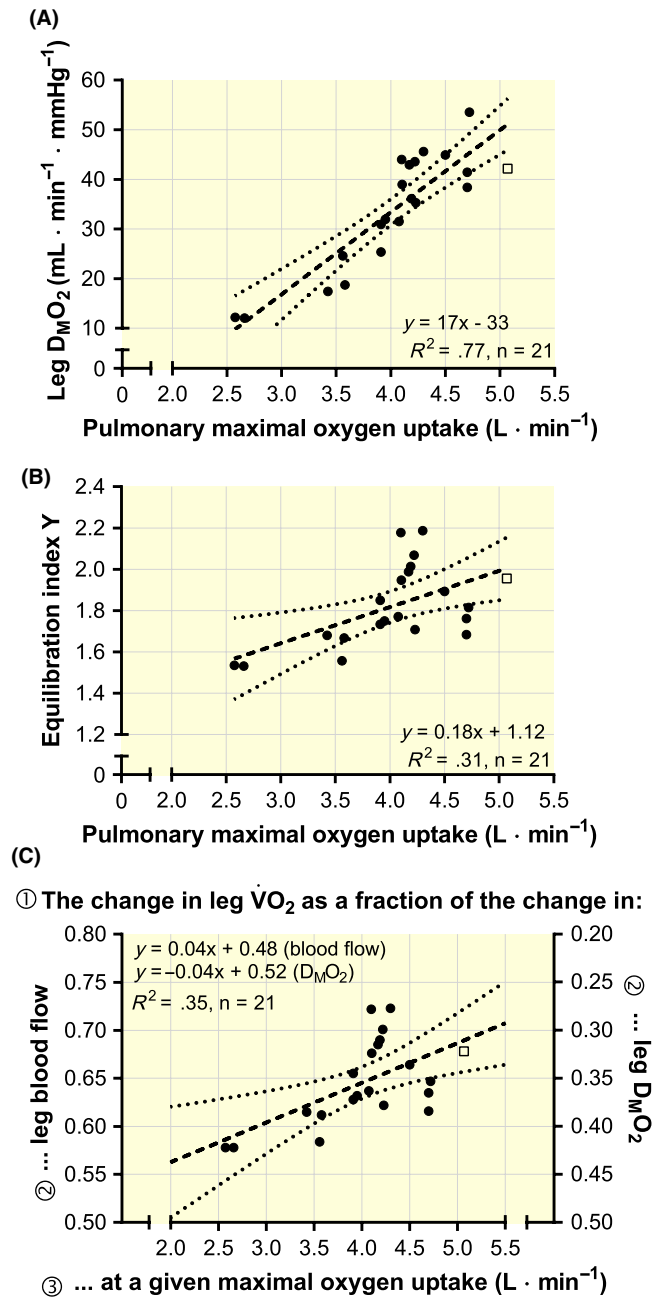


FIGURE 6 The relationship between pulmonary maximal oxygen uptake ($\dot{V}O_{2\max}$) and one-leg muscle O₂ diffusing capacity ($D_{M}O_2$; A), the equilibration index Y (B), calculated using the Piiper and Scheid model, and the fractional extent to which $\dot{V}O_{2\max}$ is expected to change if $D_{M}O_2$ or leg blood flow (LBF) is changed alone (C).²⁷ Data are mean values from studies reported in Table 2

detect actual improvements in systemic \bar{O}_2 extraction fraction when the $a\text{-}\bar{v}O_2$ difference was mostly unchanged, as endurance training may have evoked an accompanying reduction in CaO_2 . Therefore, future studies should strive to measure peripheral or systemic O₂ extraction fraction directly, or at least combine the calculations of $a\text{-}\bar{v}O_2$ difference with measurement of CaO_2 (arterial catheter). Actually, in the endurance training studies where peripheral O₂ extraction fraction

was measured directly during maximal exercise (arterial and venous catheters), the vast majority found an increased $\dot{V}O_{2\max}$ extraction fraction after training.^{12,30,45-47}

A particular case, concerning the relationship between one-leg $\dot{V}O_{2\max}$ and O_2 extraction fraction (Figure 4C) and between pulmonary $\dot{V}O_{2\max}$ and two-LBF (Figure 4D) deserves some attention (the white squares). These data were collected during combined upper- and lower-body exercise (cross-country skiing using the diagonal technique) and 6.6 $L \cdot \min^{-1}$ of \dot{Q}_{\max} was distributed to the two arms.³² Hence, when combining the locomotor blood flow (arms+legs), the data fall perfectly on the regression line between blood flow and pulmonary $\dot{V}O_{2\max}$. When redistributing LBF towards other exercising musculature, the erythrocyte capillary mean transit time (MTT) is increased. Therefore, the conditions for Hb- O_2 off-loading are improved, resulting in a slightly higher O_2 extraction fraction for a given leg $\dot{V}O_2$. The same phenomenon can be seen when adding arm cycling to ongoing leg cycling⁴⁸ or vice versa,⁴⁹ which increases the O_2 extraction fraction that compensates for some of the reduction in blood flow.

3.3 | Limitations to $\dot{V}O_{2\max}$ by O_2 delivery and O_2 extraction varies with training status

The equilibration index Y was positively correlated with $\dot{V}O_{2\max}$. Therefore, endurance training leads to a situation where the muscles become gradually more O_2 -delivery limited. Thus, individuals with the highest $\dot{V}O_{2\max}$ can only achieve a further substantial improvement in $\dot{V}O_{2\max}$ by increasing O_2 delivery, a conclusion supported by the extremely low levels of Cv_fO_2 and $C\bar{v}O_2$ in these subjects. Therefore, the limiting factors to $\dot{V}O_{2\max}$ change with training status and $\dot{V}O_{2\max}$: (a) untrained,

but healthy individuals display mixed perfusion-diffusion limitations; and (b) this diffusional limitation reduces as $\dot{V}O_{2\max}$ is increased.²⁶ These conclusions are similar to those of Gifford et al,²⁵ who found a clear relationship between OXPHOS measured in permeabilized muscle fibres ex vivo and $\dot{V}O_{2\max}$ in untrained but not in trained individuals.

3.4 | Why is not all the O_2 extracted from the blood?

The entire \dot{Q}_{\max} cannot be directed to the skeletal muscles during exercise. Other organs like the brain, heart, splanchnic organs and skin need perfusion and O_2 delivery to maintain homeostasis. \dot{Q}_{\max} must also serve the O_2 demand of the respiratory muscles and the muscles in the trunk and the arms that stabilize the subject's position on the cycle ergometer, and these tissues are characterized by a substantially lower O_2 extraction than the legs during maximal exercise.^{5,50} As a mean of those investigations measuring \dot{Q}_{\max} and LBF simultaneously (Table 3), the non-leg blood flow was 6.4 $L \cdot \min^{-1}$ and was unaffected by the level of \dot{Q}_{\max} ($y = 0.002x + 6.4$; $R^2 < .001$; $n = 12$; $P > .999$).^{4,5,9,11,31,51-57} The O_2 extraction was calculated to be 68% on average for all non-leg tissues (head, trunk and arms), explaining why the \bar{O}_2 extraction fraction of the central circulation was slightly lower than in the legs (79% vs 84%, respectively; Table 3). A mean difference of 5 percentage points might be a small underestimation since the studies using right heart catheterization^{4,9,28,29,51,55} combined with arterial and femoral venous catheters indicated a mean difference of 8 percentage points. A difference of 5%-8% points fits well, since the O_2 extraction fraction of the arms, myocardium, brain and trunk range from 40% to 80% during exercise.^{5,50,58-60} Therefore, the $C\bar{v}O_2$ can never reach the same level as the Cv_fO_2 during exercise involving

TABLE 3 Data from studies measuring pulmonary O_2 uptake, cardiac output (indicator-dilution, Fick method or transpulmonary thermodilution), leg blood flow (thermodilution) and leg arteriovenous O_2 difference (a- vO_2 difference; catheters) simultaneously during maximal exercise. From these measurements, O_2 extraction fraction was calculated for the central circulation and the non-leg tissue (combined trunk, arms and head)

	Central circulation (mean \pm SD)	Two-leg circulation (mean \pm SD)	Non-leg tissue circulation (mean \pm SD)
Blood flow ($L \cdot \min^{-1}$)	25.0 \pm 2.4	18.6 \pm 3.0	6.4 \pm 1.7
Arterial O_2 content ($mL \cdot L^{-1}$)	203 \pm 10	203 \pm 10	203 \pm 10
O_2 delivery ($L \cdot \min^{-1}$)	5.03 \pm 0.60	3.77 \pm 0.63	1.26 \pm 0.32
O_2 uptake ($L \cdot \min^{-1}$)	4.02 \pm 0.65	3.19 \pm 0.65	0.83 \pm 0.24
a- vO_2 difference ($mL \cdot L^{-1}$)	160 \pm 17	172 \pm 14	137 \pm 48
O_2 extraction fraction (%)	79 \pm 8	84 \pm 5	68 \pm 26
Venous O_2 content ($mL \cdot L^{-1}$)	42 \pm 18	31 \pm 10	66 \pm 52
O_2 delivery not utilized ($L \cdot \min^{-1}$)	1.01 \pm 0.36	0.58 \pm 0.07	0.43 \pm 0.32

Note: $n = 12$ (articles)^{4,5,9,11,31,51-57} or $n = 117$ (subjects).

the legs and was calculated to reach a minimum of $\sim 15 \text{ mL} \cdot \text{L}^{-1}$ in subjects having a $\dot{V}O_{2\text{max}}$ of $6 \text{ L} \cdot \text{min}^{-1}$ (Figure 1H). To our knowledge, the lowest $\text{C}\bar{V}O_2$ measured at sea level using right heart (atrium) catheterization is $20.1 \text{ mL} \cdot \text{L}^{-1}$ (group mean) in athletes with a $\dot{V}O_{2\text{max}}$ of $5.1 \text{ L} \cdot \text{min}^{-1}$.²⁹ A slightly lower value was measured in one of these cross-country skiers ($15.5 \text{ mL} \cdot \text{L}^{-1}$), and a mean value of $18.6 \text{ mL} \cdot \text{L}^{-1}$ has been measured in moderately trained individuals after acclimatizing to 6500 metres above sea level⁶¹; indicating that $15 \text{ mL} \cdot \text{L}^{-1}$ or lower is approachable.

The highest recorded leg O_2 extraction fraction was 93% (group mean)²⁹ and the regression models indicated a plateau at $\sim 95\%$ within physiological limits for pulmonary $\dot{V}O_{2\text{max}}$. Hence, a minimum of $\sim 10 \text{ mL } O_2$ remains in each litre of femoral venous blood associated with a PO_2 of $\sim 10 \text{ mmHg}$, even for the best trained individuals. In this situation, a PO_2 gradient persists between the blood and myoglobin (myoglobin/intracellular PO_2 : $\sim 1\text{-}2 \text{ mmHg}$),⁶² where myoglobin-facilitated diffusion should proceed given the high myoglobin O_2 affinity (myoglobin P_{50O_2} : $\sim 5 \text{ mmHg}$) and the low myoglobin SO_2 at maximal exercise.⁶² However, according to the Fick law of diffusion, the diffusive flux is directly proportional to the PO_2 gradient and will, thus, gradually decrease along the capillary and be very small when approaching low capillary PO_2 values such as 10 mmHg . It has also been shown that the primary site of resistance to O_2 diffusion is between the capillaries and the sarcoplasm and it has been estimated that the “critical capillary PO_2 ” needed to overcome this resistance may be as high as $10\text{-}20 \text{ mmHg}$.⁶²⁻⁶⁵ The remaining O_2 may, therefore, represent diffusional limitations across the combined capillary wall, interstitium and sarcolemma barriers together with a MTT that is too short for complete Hb- O_2 off-loading. This is supported by the need for an infinitesimal PO_2 gradient for O_2 to diffuse from the sarcoplasm to cytochrome c oxidase⁶⁶ and the estimate that a mitochondrial PO_2 of $\sim 1 \text{ mmHg}$ may be sufficient to support maximal mitochondrial respiration.^{67,68} The remaining O_2 may also represent muscle metabolism-perfusion mismatch^{69,70} and an inevitable lower O_2 extraction from the blood perfusing the skin, connective tissue, fat and bone marrow of the leg causing venous admixture. In this context, the end-capillary PO_2 , assessed using video microscopy, was found to be lower than the PO_2 both in the venule (O_2 micro-electrode) and vein (blood gas) draining the muscle region of interest.⁷¹ Hence, the lowest femoral venous PO_2 values of $\sim 10 \text{ mmHg}$ indicates an even lower end-capillary PO_2 in the capillaries adjacent to the most metabolically active muscle regions during maximal exercise, possibly approaching $\sim 5 \text{ mmHg}$. Therefore, no matter which kind of limitation prevails, it is highly unlikely that leg O_2 extraction fraction can improve much further, and that a theoretical threshold of $\sim 95\%$ exists because of the above diffusional and distributional limitations and barriers.

4 | THE MECHANISMS EXPLAINING THE IMPROVEMENTS OF O_2 EXTRACTION WITH TRAINING

The systemic \bar{O}_2 extraction fraction may increase through two main mechanisms with training: (a) by directing a higher fraction of \dot{Q}_{max} to the exercising muscles and (b) by increasing the peripheral O_2 extraction fraction.

Both in trained and untrained subjects, during exercise with a large muscle mass (such as running and cycling), the muscle-specific blood flow (per unit of mass) is restrained as a result of sympathetically mediated vasoconstriction of peripheral vascular beds, caused by a limited \dot{Q}_{max} .^{9,32,48} Even in “untrained” leg skeletal muscle, the reserve in vasodilatory capacity is very high and supports 2-3 times larger blood flow per unit of mass, as observed during dynamic one-legged knee extension.⁷² Simply increasing \dot{Q}_{max} (for instance, by training), without any peripheral adaptations, may increase the systemic \bar{O}_2 extraction fraction by two mechanisms. First, the recruitment of a larger portion of the already existing capillary network may reduce diffusion distances and thereby increase the O_2 extraction. This additional recruitment may also serve to maintain MTT despite increased LBF. Second, a larger fraction of \dot{Q}_{max} will flow through the exercising muscles (Figure 7) because the non-exercising tissue blood flow is independent of \dot{Q}_{max} in healthy young subjects (at $\sim 6.4 \text{ L} \cdot \text{min}^{-1}$, see section 3.4).^{4,5,9,11,31,51-57} Consequently, even without any peripheral adaptations, the systemic \bar{O}_2 extraction fraction may increase when \dot{Q}_{max} and LBF are elevated with training.

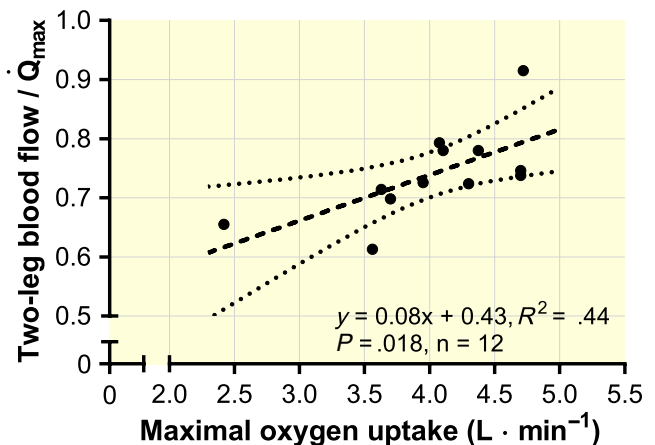


FIGURE 7 The fraction of maximal cardiac output (\dot{Q}_{max}) that is directed to the legs during maximal exercise (cycling) as a function of $\dot{V}O_{2\text{max}}$. The included studies measured \dot{Q}_{max} by using the indicator-dilution method, Fick method or transpulmonary thermodilution, and leg blood flow was measured by thermodilution.^{4,5,9,11,31,51-57} Note that the uppermost data point (0.915; ie, only $2.2 \text{ L} \cdot \text{min}^{-1}$ in calculated non-leg blood flow) is supra-physiological, but the correlation was similar after its exclusion ($R^2 = .42$)

The peripheral O_2 extraction depends on the interplay between several factors: (a) the kinetics of O_2 off-loading from Hb; (b) the erythrocyte MTT, which is determined by the blood flow, the capillary density, the capillary recruitment and the degree of matching of blood flow distribution to the metabolic demand; (c) the diffusional O_2 conductance over the combined capillary wall, interstitium and sarcolemma barriers; and (d) the muscle oxidative capacity, the mitochondrial $p50$ and the mitochondrial activation.^{10,29,73}

A right-shifted O_2 -Hb dissociation curve (elevated P_{50O_2}) increases the O_2 extraction fraction in pump-perfused dog muscle.⁷⁴ A close relationship has also been demonstrated between O_2 extraction fraction and in vivo P_{50O_2} in humans during exercise.²⁹ Very few of the studies included in the present analysis reported the in vivo P_{50O_2} , but it was possible to calculate it from the other blood gas parameters using Kelman's Equation⁷⁵ after assuming a femoral venous blood temperature of 39.0°C at maximal exercise.^{9,55,76} Based on 15 of the studies presented in Table 2, the P_{50O_2} was linearly associated with leg O_2 extraction fraction ($R^2 = .27$; $n = 15$; $P = .048$). Despite this relationship, a high P_{50O_2} does not seem to be compulsory to achieve high O_2 extraction during whole-body maximal exercise, as demonstrated in experiments using a small dose of carbon monoxide (carboxyhaemoglobin at 6%-7%), which left-shifts the ODC without a negative impact on O_2 extraction fraction.⁵⁶

Increased MTT has the potential to increase O_2 extraction, but whether this occurs after endurance training is determined by the balance between the changes in blood flow and the capillary blood volume. Capillary density typically improves by 10-30% after 4-24 weeks of endurance training,⁷⁷⁻⁷⁹ which is similar to the changes in $\dot{V}O_{2max}$ for this training duration.⁷⁸⁻⁸⁰ Moreover, cross-sectional data indicate a similar difference in capillary density to that of $\dot{V}O_{2max}$ between untrained and endurance trained men.⁴¹ Therefore, the capillary growth probably maintains the MTT despite elevated \dot{Q}_{max} and peripheral blood flow after training. In support, similar improvements in arm blood flow and capillary density have been observed after a period of arm training, causing no change in the calculated MTT.⁴⁷ The arm O_2 extraction fraction was increased in the same study, suggesting that elevated MTT is not the primary mechanism by which O_2 extraction is improved after training. However, this may differ between arms and legs (ie, small vs large muscle mass exercise). Moreover, in the calculation of MTT in the study mentioned above, full capillary recruitment was assumed. Therefore, even though the changes in capillary density and muscle blood flow share magnitudes after endurance training, the MTT may still be increased if the capillary recruitment is altered.

An increased capillary-to-fibre ratio after endurance training increases the number of contact points between the capillary and the muscle fibre. This increases the

diffusional surface area that, according to the Fick law of diffusion, increases the diffusive flux in a directly proportional manner. Therefore, the capillary-to-fibre ratio is regarded as a critical determinant of O_2 diffusion from the erythrocytes to the cytoplasm.^{81,82} As an example, a larger diffusional area and shorter diffusional distance are proposed to contribute to the higher O_2 extraction fraction in the legs than in the arms during exercise.²⁹ Moreover, if the capillary recruitment is changed with training, this may also affect the effective diffusional surface area similarly to de novo capillarization.

During whole-body maximal exercise, the oxidative capacity of skeletal muscle exceeds the O_2 delivery, as illustrated by the twofold higher $\dot{V}O_2$ per unit of muscle mass during dynamic one-legged knee extension compared to cycling exercise (approximately 2.5 vs 20 kg active muscle mass, respectively).^{10,72} Therefore, the leg muscles possess an oxidative reserve capacity at $\dot{V}O_{2max}$ during whole-body exercise, which has frequently been used as an argument to indicate that the large improvements in mitochondrial and capillary networks after endurance training are likely only crucial for improvements in endurance performance and do not affect the limiting factors to $\dot{V}O_{2max}$.⁸³ In support of this view, the calculated \bar{O}_2 extraction fraction is maintained or increases after prolonged bed rest (3-6 weeks), although a substantial reduction in mitochondrial volume density occurs.^{84,85} However, the \bar{O}_2 extraction fraction depends on the interactions between several factors. For instance, by acutely decreasing \dot{Q}_{max} and LBF using β -adrenergic blockade, $a-\bar{v}O_2$ difference and $a-v_fO_2$ difference increase during submaximal and maximal exercise, facilitated by increased erythrocyte MTT.^{86,87} This is substantiated by the positive relationship between the ratio of OXPHOS/ O_2 delivery and the leg O_2 extraction fraction,¹⁰ meaning that the balance between muscle oxidative capacity and blood flow (ie, oxidative capacity and MTT) is more critical for O_2 extraction than any of these factors alone. Therefore, as bed rest reduces \dot{Q}_{max} dramatically but causes only a minor change in capillary density,^{84,85} the MTT is elevated, and the ratio of OXPHOS/ O_2 delivery is probably the same, in favour of increased or maintained \bar{O}_2 extraction fraction. In contrast, by changing the exercise mode from upright to supine cycling after bed rest, which preserves \dot{Q}_{max} at the pre-bed rest level, the calculated $a-\bar{v}O_2$ difference is decreased (154 to 120 mL · L⁻¹).⁸⁸ Similarly, after a dog gastrocnemius muscle was immobilized for 3 weeks, followed by electrical stimulation to $\dot{V}O_{2max}$ while being pump perfused to receive a similar O_2 delivery as a control muscle, the O_2 extraction fraction was dramatically reduced.⁸² Therefore, muscle oxidative capacity seems to play a role in determining O_2 extraction, and the bed rest studies need to be evaluated carefully because of the consequences for peripheral MTT.

If \bar{O}_2 extraction fraction improves after endurance training, is probably affected by the balance between central and peripheral adaptations. For instance, after 2 weeks of high-intensity interval training that elevated the cytochrome c oxidase activity by 20% but caused no change in \dot{Q}_{\max} , $\dot{V}O_{2\max}$ was increased by 8% and was entirely attributed to the improved systemic (calculated $a-\bar{v}O_2$ difference) and leg (increased deoxyhaemoglobin and decreased tissue oxygenation index in Vastus Lateralis, assessed using NIRS) O_2 extraction.⁸⁹ However, after 3-8 weeks of endurance training, improvements in \dot{Q}_{\max} explain almost the entire increase in $\dot{V}O_{2\max}$, as indicated by meta-regression.⁴⁴ If the training lasts longer (>8 weeks), enhancements of \dot{Q}_{\max} decelerate and improvements in $a-\bar{v}O_2$ difference are again evident.^{44,90} Therefore, the peripheral adaptations are probably just sufficient to counteract the “negative influence” of elevated \dot{Q}_{\max} and LBF on MTT in periods with large central adaptations, and improvements in \bar{O}_2 extraction fraction is likely only evident when the peripheral adaptations largely surpass those of the central circulation. This can be substantiated by findings from one-legged endurance training that induces robust peripheral adaptations without stimulating the central circulation substantially and commonly improves leg $a-v_fO_2$ difference by 5-10 mL · L⁻¹.^{30,45}

The mitochondrial volume density can differ by as much as 150% between untrained and well-trained individuals in extreme cases (eg, ~4 vs ~10 vol. %) ^{91,92} and can improve by as much as ~40%-55% after 6 weeks of endurance training in previously sedentary individuals.^{38,93,94} Why does this disproportionate adaptation occur when the muscle already possesses an oxidative reserve capacity? Does it have any physiological meaning for $\dot{V}O_{2\max}$ or is it only important for improvements in, for example, fat oxidation⁹⁵ and the lactate threshold,⁹⁶ thus improving endurance?

Although an impressive increase in leg O_2 extraction fraction from 72% to 82% has been reported after only 9 weeks of intense endurance training in previously sedentary subjects,¹² we propose that remarkable increases in muscle oxidative capacity are needed to achieve the outstanding leg O_2 extraction fraction observed in elite athletes (close to 95%).^{29,97} By analogy, the oxidative reserve capacity may act as a “bottomless pit”, keeping the myoglobin SO_2 and intracellular PO_2 low. This, in turn, maintains the PO_2 gradient between the capillary and the muscle cell, promoting O_2 diffusion and O_2 extraction even at a very low capillary PO_2 .

Emerging evidence suggests that the mitochondrial volume density is increased while their intrinsic OXPHOS (OXPHOS divided by mitochondrial volume density or citrate synthase activity) is unchanged^{89,98,99} and sometimes even reduced^{94,100} after training. Since the mitochondrial

respiratory rate and the ex vivo mitochondrial p50 increase in parallel,^{10,73} the unchanged or reduced intrinsic OXPHOS after training may permit an increased OXPHOS per unit of muscle mass while preserving (or increasing) the mitochondrial O_2 affinity (ie, by keeping the mitochondrial p50 low).⁷³ Thus, a large pool of mitochondria with high O_2 affinity may preserve mitochondrial activation at low O_2 availability (low capillary PO_2) and promote peripheral O_2 extraction, but is yet to be experimentally tested. Moreover, the subsarcolemmal mitochondrial population increases relatively more than the intermyofibrillar population after endurance training.^{93,94} These mitochondrial clusters in close proximity to the capillaries may, speculatively, amplify the O_2 concentration gradient, shorten the diffusional distance and, thus, promote O_2 diffusion across the sarcolemma¹⁰¹ and enable further O_2 extraction at the end of the capillaries.

As shown in Figure 6C, a subject's $\dot{V}O_{2\max}$ becomes gradually less sensitive to adaptations improving diffusion when $\dot{V}O_{2\max}$ is already high. Therefore, to raise the O_2 extraction fraction even slightly (eg, 2%), it is likely that more substantial improvement in peripheral adaptations is needed. However, a change in leg O_2 extraction fraction from, for example, 93% to 95% would only have a small impact on whole-body $\dot{V}O_{2\max}$: for an athlete with a $\dot{V}O_{2\max}$ of 5 L · min⁻¹, a two-LBF of 24 L · min⁻¹ (\dot{Q}_{\max} : ~31 L · min⁻¹) and an CaO_2 of 190 mL · L⁻¹, the $\dot{V}O_{2\max}$ would only increase by ~90 mL · min⁻¹ (1.8%). In comparison, an increase of 1 L · min⁻¹ in two-LBF would increase $\dot{V}O_{2\max}$ by ~170 mL · min⁻¹ (3.4%) if all other factors remained the same.

5 | STUDY CONSIDERATIONS

The data were collected from several research groups and published over six decades (1958-2017) using a variety of gas analysers, flow sensors, methods to determine blood O_2 content and PO_2 , and several procedures to analyse the indicator-dilution and blood temperature curves for \dot{Q}_{\max} and LBF measurements respectively. Therefore, for a given $\dot{V}O_{2\max}$, the between-subject variability presented here may be overestimated. Moreover, several different averaging strategies for $\dot{V}O_2$ and the associated variables have likely been applied (rarely stated in the manuscripts). Despite these potential sources of noise, in general, the studies' mean values converged to similar values. The fact that, despite the combination of several measurements with distinct methods (such as pulmonary gas exchange, thermodilution and blood gas analyses), the integrations of the obtained values fitted into the physiological range and agreed between studies, demonstrates the quality of these studies and the robustness of the analysis presented here.

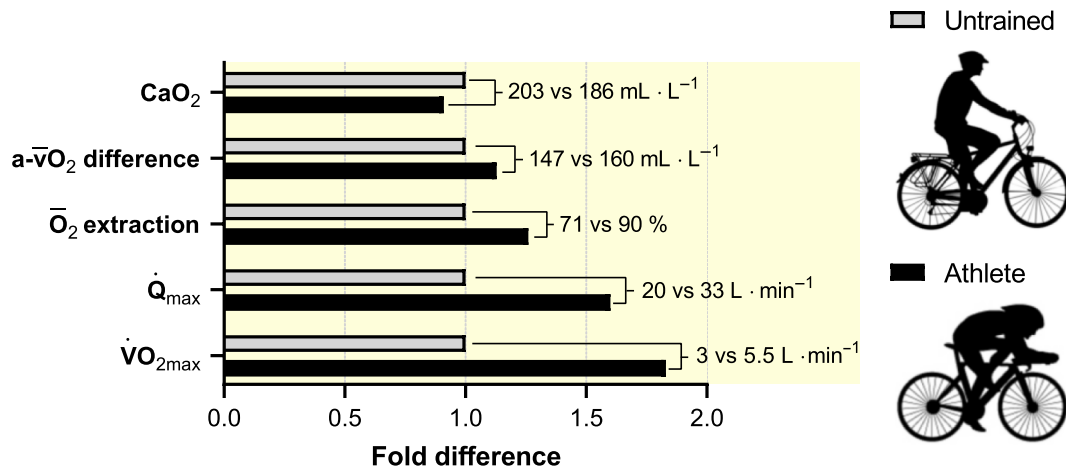


FIGURE 8 A comparison of an untrained individual and an elite endurance athlete with maximal oxygen uptakes ($\dot{V}O_{2\max}$) of 3.0 and 5.5 L · min⁻¹ respectively. The maximal cardiac output (\dot{Q}_{\max}), systemic O₂ extraction fraction (\bar{O}_2 extraction), arterial to mixed venous O₂ difference (a- \bar{v} O₂ difference) and arterial O₂ content (CaO₂) were calculated using the regression equations presented in Figure 1

6 | CONCLUSION AND PERSPECTIVE

In conclusion, measurements of \dot{Q}_{\max} and LBF show that O₂ delivery is the primary determinant of whole-body and limb $\dot{V}O_{2\max}$. However, we also show that a very high O₂ extraction fraction contributes to the remarkably high $\dot{V}O_{2\max}$ in well-trained individuals and elite endurance athletes. To reinforce this conclusion we can, using the regression lines established in the present investigation, compare a typically sedentary subject and an elite endurance athlete with a large difference in $\dot{V}O_{2\max}$ (3.0 vs 5.5 L · min⁻¹): the elite athlete has a 1.83-fold higher $\dot{V}O_{2\max}$, a 1.60-fold higher \dot{Q}_{\max} and a 1.26-fold higher \bar{O}_2 extraction fraction (Figure 8). However, because of the lower CaO₂, the a- \bar{v} O₂ difference is only 1.13-fold higher in the elite athlete. This also stresses that a- \bar{v} O₂ difference and \bar{O}_2 extraction fraction cannot be used interchangeably when evaluating central versus peripheral limitations to $\dot{V}O_{2\max}$. Finally, the limitations for whole-body $\dot{V}O_{2\max}$ change with training status, with an accentuated O₂ delivery limitation and conversely a decreasing O₂ diffusional limitation with increasing $\dot{V}O_{2\max}$.

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CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

AUTHOR CONTRIBUTIONS

Conception and design of the investigation: ØS, JH, CC, JALC. Literature search and analysis of data: ØS. Interpretation of data: ØS, JH, CC, JALC, BR. Writing the

first draft of the manuscript: ØS. Revising and approving the final version: ØS, JH, CC, JALC, BR.

DATA AVAILABILITY STATEMENT

Data sharing is not applicable to this article, as no new data were created in this study.

ORCID

Øyvind Skattebo  <https://orcid.org/0000-0003-0771-9715>
 Jose A. L. Calbet  <https://orcid.org/0000-0002-9215-6234>
 Bjarne Rud  <https://orcid.org/0000-0001-9692-6542>
 Carlo Capelli  <https://orcid.org/0000-0002-3278-1337>
 Jostein Hallén  <https://orcid.org/0000-0002-6646-0734>

REFERENCES

- Joyner MJ, Casey DP. Regulation of increased blood flow (hyperemia) to muscles during exercise: a hierarchy of competing physiological needs. *Physiol Rev.* 2015;95(2):549-601.
- Saltin B, Åstrand PO. Maximal oxygen uptake in athletes. *J Appl Physiol.* 1967;23(3):353-358.
- Bouchard C, Sarzynski MA, Rice TK, et al. Genomic predictors of the maximal O₂ uptake response to standardized exercise training programs. *J Appl Physiol (1985).* 2011;110(5):1160-1170.
- Mortensen SP, Dawson EA, Yoshiga CC, et al. Limitations to systemic and locomotor limb muscle oxygen delivery and uptake during maximal exercise in humans. *J Physiol.* 2005;566(Pt 1):273-285.
- Calbet JAL, Gonzalez-Alonso J, Helge JW, et al. Cardiac output and leg and arm blood flow during incremental exercise to exhaustion on the cycle ergometer. *J Appl Physiol (1985).* 2007;103(3):969-978.
- di Prampero PE, Ferretti G. Factors limiting maximal oxygen consumption in humans. *Respir Physiol.* 1990;80(2-3):113-127.
- Saltin B, Calbet JA. Point: in health and in a normoxic environment, VO₂ max is limited primarily by cardiac output and locomotor muscle blood flow. *J Appl Physiol (1985).* 2006;100(2):744-745.

8. Wagner PD. Counterpoint: In health and in a normoxic environment, VO_2 max is not limited primarily by cardiac output and locomotor muscle blood flow. *J Appl Physiol* (1985). 2006;100(2):745-747.
9. Mortensen SP, Damsgaard R, Dawson EA, Secher NH, Gonzalez-Alonso J. Restrictions in systemic and locomotor skeletal muscle perfusion, oxygen supply and VO_2 during high-intensity whole-body exercise in humans. *J Physiol*. 2008;586(10):2621-2635.
10. Cardinale DA, Larsen FJ, Jensen-Urstad M, et al. Muscle mass and inspired oxygen influence oxygen extraction at maximal exercise: role of mitochondrial oxygen affinity. *Acta Physiol*. 2019;225(1):e13110.
11. Gonzalez-Alonso J, Calbet JA. Reductions in systemic and skeletal muscle blood flow and oxygen delivery limit maximal aerobic capacity in humans. *Circulation*. 2003;107(6):824-830.
12. Roca J, Agusti AG, Alonso A, et al. Effects of training on muscle O_2 transport at $\text{VO}_{2\text{max}}$. *J Appl Physiol* (1985). 1992;73(3):1067-1076.
13. Lundby C, Robach P. Performance enhancement: what are the physiological limits? *Physiology*. 2015;30(4):282-292.
14. Lundby C, Montero D, Joyner M. Biology of $\text{VO}_{2\text{max}}$: looking under the physiology lamp. *Acta Physiol*. 2017;220(2):218-228.
15. Levine BD. $\text{VO}_{2\text{max}}$: what do we know, and what do we still need to know? *J Physiol*. 2008;586(1):25-34.
16. Ekblom B, Hermansen L. Cardiac output in athletes. *J Appl Physiol*. 1968;25(5):619-625.
17. Ekblom B, Åstrand PO, Saltin B, Stenberg J, Wallstrom B. Effect of training on circulatory response to exercise. *J Appl Physiol*. 1968;24(4):518-528.
18. Sawka MN, Convertino VA, Eichner ER, Schnieder SM, Young AJ. Blood volume: importance and adaptations to exercise training, environmental stresses, and trauma/sickness. *Med Sci Sports Exerc*. 2000;32(2):332-348.
19. Powers SK, Lawler J, Dempsey JA, Dodd S, Landry G. Effects of incomplete pulmonary gas exchange on $\text{VO}_{2\text{max}}$. *J Appl Physiol* (1985). 1989;66(6):2491-2495.
20. Nielsen HB. Arterial desaturation during exercise in man: implication for O_2 uptake and work capacity. *Scand J Med Sci Sports*. 2003;13(6):339-358.
21. Hermansen L, Ekblom B, Saltin B. Cardiac output during submaximal and maximal treadmill and bicycle exercise. *J Appl Physiol*. 1970;29(1):82-86.
22. Gleser MA. Effects of hypoxia and physical training on hemodynamic adjustments to one-legged exercise. *J Appl Physiol*. 1973;34(5):655-659.
23. Mitchell JH, Sproule BJ, Chapman CB. The physiological meaning of the maximal oxygen intake test. *J Clin Invest*. 1958;37(4):538-547.
24. Åstrand PO, Cuddy TE, Saltin B, Stenberg J. Cardiac output during submaximal and maximal work. *J Appl Physiol*. 1964;19:268-274.
25. Gifford JR, Garten RS, Nelson AD, et al. Symmorphosis and skeletal muscle $\text{VO}_{2\text{max}}$. In vivo and in vitro measures reveal differing constraints in the exercise-trained and untrained human. *J Physiol*. 2016;594(6):1741-1751.
26. Wagner PD. Systemic oxygen transport and utilization. *J Breath Res*. 2008;2(2):024001.
27. Piiper J. Perfusion, diffusion and their heterogeneities limiting blood-tissue O_2 transfer in muscle. *Acta Physiol Scand*. 2000;168(4):603-607.
28. Roca J, Hogan MC, Story D, et al. Evidence for tissue diffusion limitation of $\text{VO}_{2\text{max}}$ in normal humans. *J Appl Physiol* (1985). 1989;67(1):291-299.
29. Calbet JA, Holmberg HC, Rosdahl H, van Hall G, Jensen-Urstad M, Saltin B. Why do arms extract less oxygen than legs during exercise? *Am J Physiol Regul Integr Comp Physiol*. 2005;289(5):R1448-R1458.
30. Klausen K, Secher NH, Clausen JP, Hartling O, Trap-Jensen J. Central and regional circulatory adaptations to one-leg training. *J Appl Physiol*. 1982;52(4):976-983.
31. Calbet JAL, González-Alonso J, Helge JW, et al. Central and peripheral hemodynamics in exercising humans: leg vs arm exercise. *Scand J Med Sci Sports*. 2015;25(Suppl 4):144-157.
32. Calbet JA, Jensen-Urstad M, van Hall G, Holmberg HC, Rosdahl H, Saltin B. Maximal muscular vascular conductances during whole body upright exercise in humans. *J Physiol*. 2004;558(Pt 1):319-331.
33. Scharhag J, Schneider G, Urhausen A, Rochette V, Kramann B, Kindermann W. Athlete's heart: right and left ventricular mass and function in male endurance athletes and untrained individuals determined by magnetic resonance imaging. *J Am Coll Cardiol*. 2002;40(10):1856-1863.
34. Skattebo Ø, Bjerring AW, Auensen M, et al. Blood volume expansion does not explain the increase in peak oxygen uptake induced by 10 weeks of endurance training. *Eur J Appl Physiol*. 2020;120(5):985-999.
35. Levine BD, Lane LD, Buckley JC, Friedman DB, Blomqvist CG. Left ventricular pressure-volume and Frank-Starling relations in endurance athletes. Implications for orthostatic tolerance and exercise performance. *Circulation*. 1991;84(3):1016-1023.
36. Arbab-Zadeh A, Perhonen M, Howden E, et al. Cardiac remodeling in response to 1 year of intensive endurance training. *Circulation*. 2014;130(24):2152-2161.
37. Heinicke K, Wolfarth B, Winchenbach P, et al. Blood volume and hemoglobin mass in elite athletes of different disciplines. *Int J Sports Med*. 2001;22(7):504-512.
38. Montero D, Cathomen A, Jacobs RA, et al. Haematological rather than skeletal muscle adaptations contribute to the increase in peak oxygen uptake induced by moderate endurance training. *J Physiol*. 2015;593(20):4677-4688.
39. Bassett DR Jr, Howley ET. Maximal oxygen uptake: "classical" versus "contemporary" viewpoints. *Med Sci Sports Exerc*. 1997;29(5):591-603.
40. Jacobs RA, Lundby C. Mitochondria express enhanced quality as well as quantity in association with aerobic fitness across recreationally active individuals up to elite athletes. *J Appl Physiol* (1985). 2013;114(3):344-350.
41. Brodal P, Ingjer F, Hermansen L. Capillary supply of skeletal muscle fibers in untrained and endurance-trained men. *Am J Physiol*. 1977;232(6):H705-H712.
42. Kanstrup IL, Ekblom B. Acute hypervolemia, cardiac performance, and aerobic power during exercise. *J Appl Physiol Respir Environ Exerc Physiol*. 1982;52(5):1186-1191.
43. Warburton DE, Gledhill N, Jamnik VK, Krip B, Card N. Induced hypervolemia, cardiac function, $\text{VO}_{2\text{max}}$, and performance of elite cyclists. *Med Sci Sports Exerc*. 1999;31(6):800-808.
44. Montero D, Diaz-Canestro C, Lundby C. Endurance training and $\text{VO}_{2\text{max}}$: role of maximal cardiac output and oxygen extraction. *Med Sci Sports Exerc*. 2015;47(10):2024-2033.

45. Rud B, Foss O, Krstrup P, Secher NH, Hallén J. One-legged endurance training: leg blood flow and oxygen extraction during cycling exercise. *Acta Physiol.* 2012;205(1):177-185.
46. Beere PA, Russell SD, Morey MC, Kitzman DW, Higginbotham MB. Aerobic exercise training can reverse age-related peripheral circulatory changes in healthy older men. *Circulation.* 1999;100(10):1085-1094.
47. Boushel R, Ara I, Gnaiger E, et al. Low-intensity training increases peak arm VO₂ by enhancing both convective and diffusive O₂ delivery. *Acta Physiol.* 2014;211(1):122-134.
48. Secher NH, Clausen JP, Klausen K, Noer I, Trap-Jensen J. Central and regional circulatory effects of adding arm exercise to leg exercise. *Acta Physiol Scand.* 1977;100(3):288-297.
49. Volianitis S, Secher NH. Arm blood flow and metabolism during arm and combined arm and leg exercise in humans. *J Physiol.* 2002;544(Pt 3):977-984.
50. Trangmar SJ, Chiesa ST, Stock CG, Kalsi KK, Secher NH, Gonzalez-Alonso J. Dehydration affects cerebral blood flow but not its metabolic rate for oxygen during maximal exercise in trained humans. *J Physiol.* 2014;592(14):3143-3160.
51. Sullivan MJ, Knight JD, Higginbotham MB, Cobb FR. Relation between central and peripheral hemodynamics during exercise in patients with chronic heart failure. Muscle blood flow is reduced with maintenance of arterial perfusion pressure. *Circulation.* 1989;80(4):769-781.
52. Calbet JA, Lundby C, Sander M, Robach P, Saltin B, Boushel R. Effects of ATP-induced leg vasodilation on VO₂ peak and leg O₂ extraction during maximal exercise in humans. *Am J Physiol Regul Integr Comp Physiol.* 2006;291(2):R447-R453.
53. Lundby C, Boushel R, Robach P, Moller K, Saltin B, Calbet JA. During hypoxic exercise some vasoconstriction is needed to match O₂ delivery with O₂ demand at the microcirculatory level. *J Physiol.* 2008;586(1):123-130.
54. Lundby C, Robach P, Boushel R, et al. Does recombinant human Epo increase exercise capacity by means other than augmenting oxygen transport? *J Appl Physiol (1985).* 2008;105(2):581-587.
55. Munch GD, Svendsen JH, Damsgaard R, Secher NH, Gonzalez-Alonso J, Mortensen SP. Maximal heart rate does not limit cardiovascular capacity in healthy humans: insight from right atrial pacing during maximal exercise. *J Physiol.* 2014;592(2):377-390.
56. Calbet JA, Losa-Reyna J, Torres-Peralta R, et al. Limitations to oxygen transport and utilization during sprint exercise in humans: evidence for a functional reserve in muscle O₂ diffusing capacity. *J Physiol.* 2015;593(20):4649-4664.
57. Calbet JA, Boushel R, Rådegran G, Sondergaard H, Wagner PD, Saltin B. Why is VO₂ max after altitude acclimatization still reduced despite normalization of arterial O₂ content? *Am J Physiol Regul Integr Comp Physiol.* 2003;284(2):R304-316.
58. Fisher JP, Hartwich D, Seifert T, et al. Cerebral perfusion, oxygenation and metabolism during exercise in young and elderly individuals. *J Physiol.* 2013;591(7):1859-1870.
59. Duncker DJ, Bache RJ. Regulation of coronary blood flow during exercise. *Physiol Rev.* 2008;88(3):1009-1086.
60. Heiss HW, Barmeyer J, Wink K, et al. Studies on the regulation of myocardial blood flow in man. I.: training effects on blood flow and metabolism of the healthy heart at rest and during standardized heavy exercise. *Basic Res Cardiol.* 1976;71(6):658-675.
61. Sutton JR, Reeves JT, Wagner PD, et al. Operation Everest II: oxygen transport during exercise at extreme simulated altitude. *J Appl Physiol (1985).* 1988;64(4):1309-1321.
62. Gayeski TE, Honig CR. Intracellular PO₂ in long axis of individual fibers in working dog gracilis muscle. *Am J Physiol.* 1988;254(6 Pt 2):H1179-H1186.
63. Wittenberg BA, Wittenberg JB. Transport of oxygen in muscle. *Annu Rev Physiol.* 1989;51:857-878.
64. Roy TK, Popel AS. Theoretical predictions of end-capillary PO₂ in muscles of athletic and nonathletic animals at VO_{2max}. *Am J Physiol.* 1996;271(2 Pt 2):H721-H737.
65. Poole DC, Jones AM. Oxygen uptake kinetics. *Compr Physiol.* 2012;2(2):933-996.
66. Clark A Jr, Clark PA, Connert RJ, Gayeski TE, Honig CR. How large is the drop in PO₂ between cytosol and mitochondrion? *Am J Physiol.* 1987;252(6 Pt 1):C583-587.
67. Gayeski TE, Connert RJ, Honig CR. Minimum intracellular PO₂ for maximum cytochrome turnover in red muscle in situ. *Am J Physiol.* 1987;252(5 Pt 2):H906-H915.
68. Nanadikar MS, Vergel Leon AM, Borowik S, et al. O₂ affects mitochondrial functionality ex vivo. *Redox Biol.* 2019;22:101152.
69. Heinonen IH, Kemppainen J, Kaskinoro K, et al. Regulation of human skeletal muscle perfusion and its heterogeneity during exercise in moderate hypoxia. *Am J Physiol Regul Integr Comp Physiol.* 2010;299(1):R72-R79.
70. Cano I, Roca J, Wagner PD. Effects of lung ventilation-perfusion and muscle metabolism-perfusion heterogeneities on maximal O₂ transport and utilization. *J Physiol.* 2015;593(8):1841-1856.
71. Stein JC, Ellis CG, Ellsworth ML. Relationship between capillary and systemic venous PO₂ during nonhypoxic and hypoxic ventilation. *Am J Physiol.* 1993;265(2 Pt 2):H537-H542.
72. Boushel R, Saltin B. Ex vivo measures of muscle mitochondrial capacity reveal quantitative limits of oxygen delivery by the circulation during exercise. *Int J Biochem Cell Biol.* 2013;45(1):68-75.
73. Larsen FJ, Schiffer TA, Zinner C, et al. Mitochondrial oxygen affinity increases after sprint interval training and is related to the improvement in peak oxygen uptake. *Acta Physiol.* 2020:e13463.
74. Richardson RS, Tagore K, Haseler LJ, Jordan M, Wagner PD. Increased VO_{2max} with right-shifted Hb-O₂ dissociation curve at a constant O₂ delivery in dog muscle in situ. *J Appl Physiol (1985).* 1998;84(3):995-1002.
75. Kelman GR. Digital computer subroutine for the conversion of oxygen tension into saturation. *J Appl Physiol.* 1966;21(4):1375-1376.
76. Trangmar SJ, Chiesa ST, Kalsi KK, Secher NH, Gonzalez-Alonso J. Whole body hyperthermia, but not skin hyperthermia, accelerates brain and locomotor limb circulatory strain and impairs exercise capacity in humans. *Physiol Rep.* 2017;5(2):e13108.
77. Hoier B, Hellsten Y. Exercise-induced capillary growth in human skeletal muscle and the dynamics of VEGF. *Microcirculation.* 2014;21(4):301-314.
78. Ingier F. Effects of endurance training on muscle fibre ATP-ase activity, capillary supply and mitochondrial content in man. *J Physiol.* 1979;294:419-432.
79. Klausen K, Andersen LB, Pelle I. Adaptive changes in work capacity, skeletal muscle capillarization and enzyme levels during training and detraining. *Acta Physiol Scand.* 1981;113(1):9-16.
80. Milanovic Z, Sporis G, Weston M. Effectiveness of high-intensity interval training (HIT) and continuous endurance training for VO_{2max} improvements: a systematic review and meta-analysis of controlled trials. *Sports Med.* 2015;45(10):1469-1481.
81. Wagner PD. Diffusive resistance to O₂ transport in muscle. *Acta Physiol Scand.* 2000;168(4):609-614.

82. Hepple RT, Hogan MC, Stary C, Bebout DE, Mathieu-Costello O, Wagner PD. Structural basis of muscle O₂ diffusing capacity: evidence from muscle function in situ. *J Appl Physiol* (1985). 2000;88(2):560-566.
83. Bassett DR Jr, Howley ET. Limiting factors for maximum oxygen uptake and determinants of endurance performance. *Med Sci Sports Exerc*. 2000;32(1):70-84.
84. Ferretti G, Antonutto G, Denis C, et al. The interplay of central and peripheral factors in limiting maximal O₂ consumption in man after prolonged bed rest. *J Physiol*. 1997;501(Pt 3):677-686.
85. Saltin B, Blomqvist G, Mitchell JH, Johnson RL Jr, Wildenthal K, Chapman CB. Response to exercise after bed rest and after training. *Circulation*. 1968;38(5 Suppl):VIII-78.
86. Ekblom B, Goldbarg AN, Kilbom A, Astrand PO. Effects of atropine and propranolol on the oxygen transport system during exercise in man. *Scand J Clin Lab Invest*. 1972;30(1):35-42.
87. Pawelczyk JA, Hanel B, Pawelczyk RA, Warberg J, Secher NH. Leg vasoconstriction during dynamic exercise with reduced cardiac output. *J Appl Physiol* (1985). 1992;73(5):1838-1846.
88. Bringard A, Pogliaghi S, Adami A, et al. Cardiovascular determinants of maximal oxygen consumption in upright and supine posture at the end of prolonged bed rest in humans. *Respir Physiol Neurobiol*. 2010;172(1-2):53-62.
89. Jacobs RA, Flück D, Bonne TC, et al. Improvements in exercise performance with high-intensity interval training coincide with an increase in skeletal muscle mitochondrial content and function. *J Appl Physiol* (1985). 2013;115(6):785-793.
90. Murias JM, Kowalchuk JM, Paterson DH. Time course and mechanisms of adaptations in cardiorespiratory fitness with endurance training in older and young men. *J Appl Physiol* (1985). 2010;108(3):621-627.
91. Lundby C, Jacobs RA. Adaptations of skeletal muscle mitochondria to exercise training. *Exp Physiol*. 2016;101(1):17-22.
92. Hoppeler H, Luthi P, Claassen H, Weibel ER, Howald H. The ultrastructure of the normal human skeletal muscle. A morphometric analysis on untrained men, women and well-trained orienteers. *Pflugers Arch*. 1973;344(3):217-232.
93. Hoppeler H, Howald H, Conley K, et al. Endurance training in humans: aerobic capacity and structure of skeletal muscle. *J Appl Physiol* (1985). 1985;59(2):320-327.
94. Meinild Lundby AK, Jacobs RA, Gehrig S, et al. Exercise training increases skeletal muscle mitochondrial volume density by enlargement of existing mitochondria and not de novo biogenesis. *Acta Physiol*. 2018;222(1):e12976.
95. Dandanell S, Meinild-Lundby A-K, Andersen AB, et al. Determinants of maximal whole-body fat oxidation in elite cross-country skiers: role of skeletal muscle mitochondria. *Scand J Med Sci Sports*. 2018;28(12):2494-2504.
96. Coyle EF, Coggan AR, Hopper MK, Walters TJ. Determinants of endurance in well-trained cyclists. *J Appl Physiol* (1985). 1988;64(6):2622-2630.
97. Knight DR, Schaffartzik W, Poole DC, Hogan MC, Bebout DE, Wagner PD. Effects of hyperoxia on maximal leg O₂ supply and utilization in men. *J Appl Physiol* (1985). 1993;75(6):2586-2594.
98. MacInnis MJ, Zacharewicz E, Martin BJ, et al. Superior mitochondrial adaptations in human skeletal muscle after interval compared to continuous single-leg cycling matched for total work. *J Physiol*. 2017;595(9):2955-2968.
99. Granata C, Oliveira RS, Little JP, Renner K, Bishop DJ. Mitochondrial adaptations to high-volume exercise training are rapidly reversed after a reduction in training volume in human skeletal muscle. *FASEB J*. 2016;30(10):3413-3423.
100. Larsen FJ, Schiffer TA, Ørtenblad N, et al. High-intensity sprint training inhibits mitochondrial respiration through aconitase inactivation. *FASEB J*. 2016;30(1):417-427.
101. Aw TY. Intracellular compartmentation of organelles and gradients of low molecular weight species. *Int Rev Cytol*. 2000;192:223-253.
102. Bender PR, Groves BM, McCullough RE, et al. Oxygen transport to exercising leg in chronic hypoxia. *J Appl Physiol* (1985). 1988;65(6):2592-2597.
103. Knight DR, Poole DC, Schaffartzik W, et al. Relationship between body and leg VO₂ during maximal cycle ergometry. *J Appl Physiol* (1985). 1992;73(3):1114-1121.
104. Cardus J, Marrades RM, Roca J, et al. Effects of F(I)O₂ on leg VO₂ during cycle ergometry in sedentary subjects. *Med Sci Sports Exerc*. 1998;30(5):697-703.
105. Harms CA, Babcock MA, McClaran SR, et al. Respiratory muscle work compromises leg blood flow during maximal exercise. *J Appl Physiol* (1985). 1997;82(5):1573-1583.
106. van Hall G, Calbet JA, Sondergaard H, Saltin B. The re-establishment of the normal blood lactate response to exercise in humans after prolonged acclimatization to altitude. *J Physiol*. 2001;536(Pt 3):963-975.
107. Lundby C, Calbet JA, van Hall G, Saltin B, Sander M. Pulmonary gas exchange at maximal exercise in Danish lowlanders during 8 wk of acclimatization to 4,100 m and in high-altitude Aymara natives. *Am J Physiol Regul Integr Comp Physiol*. 2004;287(5):R1202-R1208.
108. Stenberg J, Åstrand PO, Ekblom B, Royce J, Saltin B. Hemodynamic response to work with different muscle groups, sitting and supine. *J Appl Physiol*. 1967;22(1):61-70.
109. Saltin B, Stenberg J. Circulatory response to prolonged severe exercise. *J Appl Physiol*. 1964;19:833-838.
110. Saltin B. Circulatory response to submaximal and maximal exercise after thermal dehydration. *J Appl Physiol*. 1964;19:1125-1132.
111. Saltin B, Grover RF, Blomqvist CG, Hartley LH, Johnson RL Jr. Maximal oxygen uptake and cardiac output after two weeks at 4,300 m. *J Appl Physiol* (1985). 1968;25(3):400-409.
112. Ekblom B. Effect of physical training on circulation during prolonged severe exercise. *Acta Physiol Scand*. 1970;78(2):145-158.
113. Lundby C, Sander M, van Hall G, Saltin B, Calbet JA. Maximal exercise and muscle oxygen extraction in acclimatizing lowlanders and high altitude natives. *J Physiol*. 2006;573(Pt 2):535-547.
114. Stenberg J, Ekblom B, Messin R. Hemodynamic response to work at simulated altitude, 4,000 m. *J Appl Physiol*. 1966;21(5):1589-1594.
115. Poole DC, Schaffartzik W, Knight DR, et al. Contribution of exercising legs to the slow component of oxygen uptake kinetics in humans. *J Appl Physiol* (1985). 1991;71(4):1245-1260.
116. Blomqvist G, Saltin B, Mitchell JH, Vastagh GF. Acute effects of ethanol ingestion on the response to submaximal and maximal exercise in man. *Circulation*. 1970;42(3):463-470.
117. Epstein S, Robinson BF, Kahler RL, Braunwald E. Effects of beta-adrenergic blockade on the cardiac response to maximal and submaximal exercise in man. *J Clin Invest*. 1965;44(11):1745-1753.
118. Epstein SE, Beiser GD, Stampfer M, Robinson BF, Braunwald E. Characterization of the circulatory response to maximal upright exercise in normal subjects and patients with heart disease. *Circulation*. 1967;35(6):1049-1062.

119. Robinson BF, Epstein SE, Kahler RL, Braunwald E. Circulatory effects of acute expansion of blood volume. *Circ Res.* 1966;19(1):26-32.
120. Proctor DN, Newcomer SC, Koch DW, Le KU, MacLean DA, Leuenberger UA. Leg blood flow during submaximal cycle ergometry is not reduced in healthy older normally active men. *J Appl Physiol (1985)*. 2003;94(5):1859-1869.
121. Harms CA, Wetter TJ, McClaran SR, et al. Effects of respiratory muscle work on cardiac output and its distribution during maximal exercise. *J Appl Physiol (1985)*. 1998;85(2):609-618.
122. Sun X-G, Hansen JE, Ting H, et al. Comparison of Exercise cardiac output by the Fick principle using oxygen and carbon dioxide. *Chest.* 2000;118(3):631-640.
123. Siebenmann C, Rasmussen P, Sørensen H, et al. Cardiac output during exercise: a comparison of four methods. *Scand J Med Sci Sports.* 2015;25(1):e20-e27.
124. Nielsen HB, Boushel R, Madsen P, Secher NH. Cerebral desaturation during exercise reversed by O₂ supplementation. *Am J Physiol Heart Circ Physiol.* 1999;277(3):H1045-H1052.
125. Woodson RD, Wills RE, Lenfant C. Effect of acute and established anemia on O₂ transport at rest, submaximal and maximal work. *J Appl Physiol Respir Environ Exerc Physiol.* 1978;44(1):36-43.
126. Turner DL, Hoppeler H, Noti C, et al. Limitations to VO_{2max} in humans after blood retransfusion. *Respir Physiol.* 1993;92(3):329-341.
127. Higginbotham MB, Morris KG, Williams RS, McHale PA, Coleman RE, Cobb FR. Regulation of stroke volume during submaximal and maximal upright exercise in normal man. *Circ Res.* 1986;58(2):281-291.
128. Ekblom B, Huot R, Stein EM, Thorstensson AT. Effect of changes in arterial oxygen content on circulation and physical performance. *J Appl Physiol.* 1975;39(1):71-75.
129. Ekblom B, Wilson G, Åstrand PO. Central circulation during exercise after venesection and reinfusion of red blood cells. *J Appl Physiol.* 1976;40(3):379-383.
130. Celsing F, Nystrom J, Pihlstedt P, Werner B, Ekblom B. Effect of long-term anemia and retransfusion on central circulation during exercise. *J Appl Physiol (1985)*. 1986;61(4):1358-1362.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

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